

## 讲座一 斑马鱼基因突变技术概述

国家斑马鱼资源中心 (CZRC)

谢训卫

国家水生生物种质资源库

国家斑马鱼资源中心

[zebrafish\\_sub@ihb.ac.cn](mailto:zebrafish_sub@ihb.ac.cn)

国家水生生物种质资源库 (NABRC)

▶ 基因敲降技术 (Gene knockdown)

▶ DNA双链断裂修复 (DSB Repair) (CZRC)

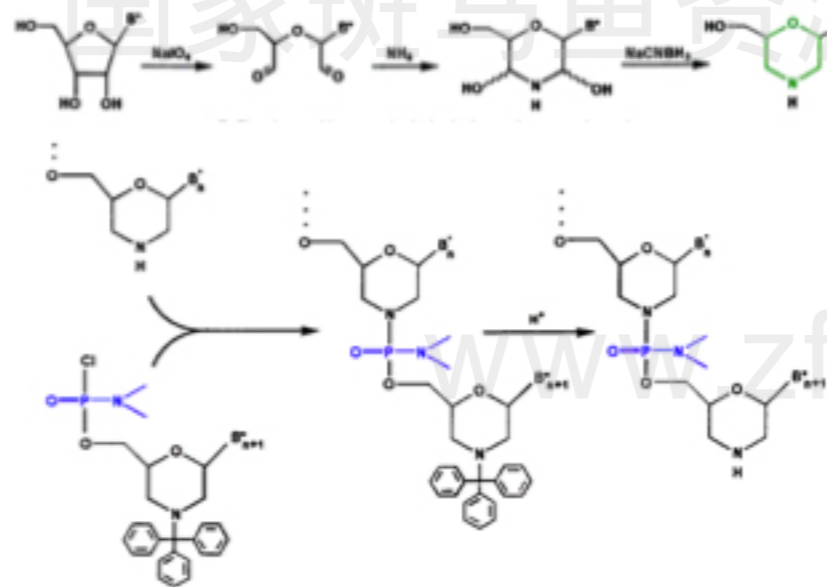
▶ 基因编辑技术 (Gene Editing)

www.zfish.cn

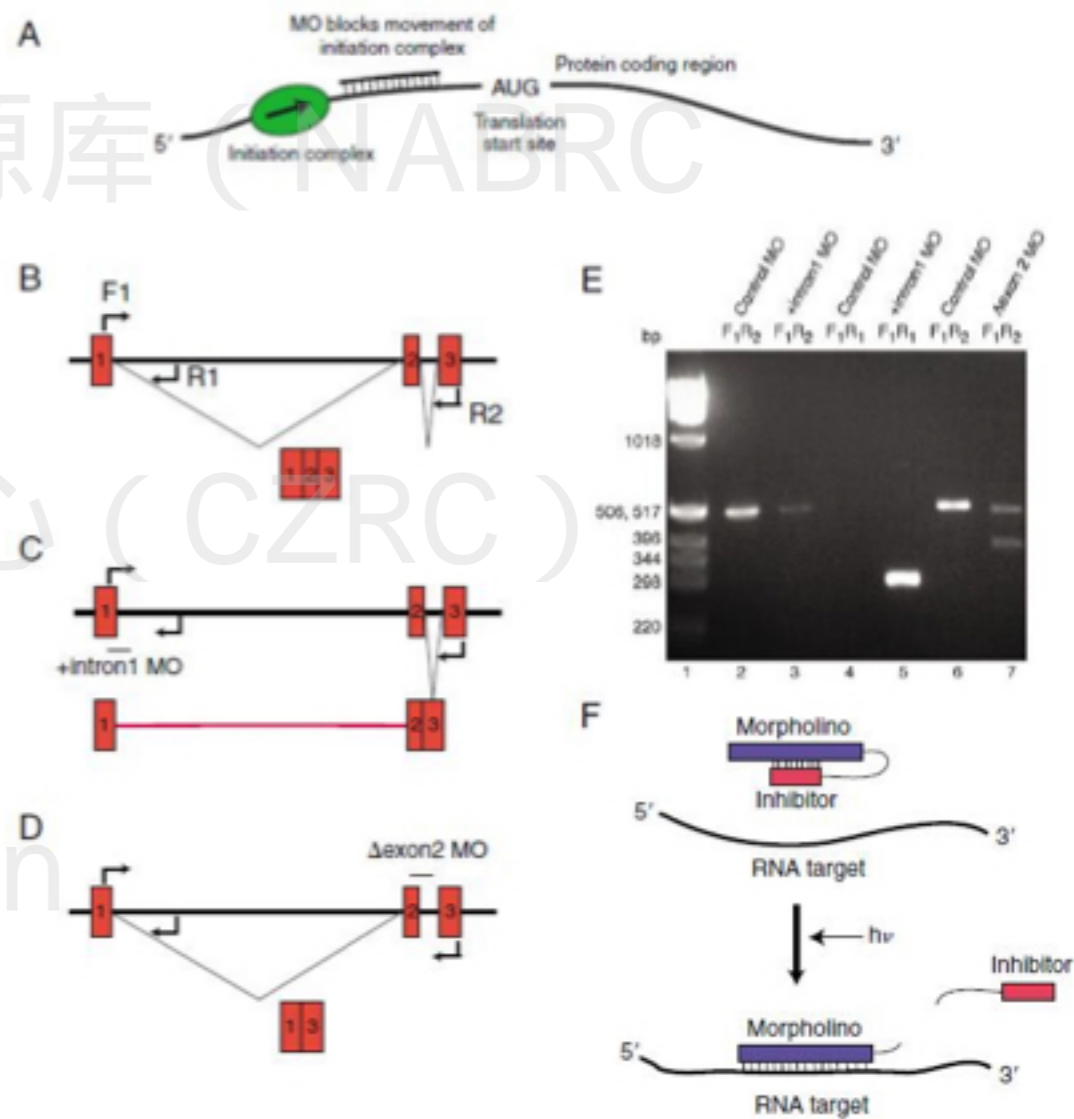
# 1、基因敲降技术

## 1.1 反义Morpholino技术:

- ① 人工合成的稳定的核酸类似物;
- ② 使用吗啉环替代核苷酸上的五碳糖环, 较**稳定**;
- ③ 寡核苷酸**单链**, ~25nt, 以标准碱基互补配对的方式与mRNA结合。
- ④ ATG-MO: 阻止翻译, 靶点位于mRNA的5' UTR 或者靠近ATG的编码区;
- ⑤ Splice-MO: 阻止剪切, 靶点位于外显子与内含子的交界处, 影响正常剪接。



Morpholino转变及组装示意图 (gene-tools公司: <http://www.gene-tools.com/>)

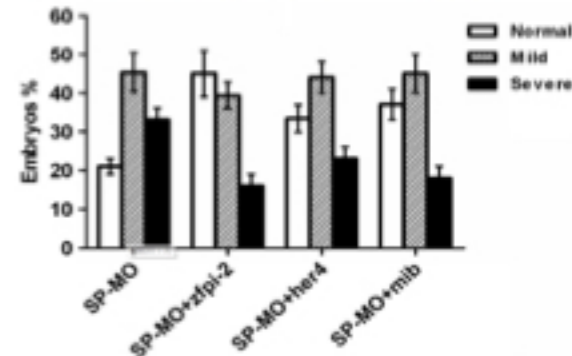
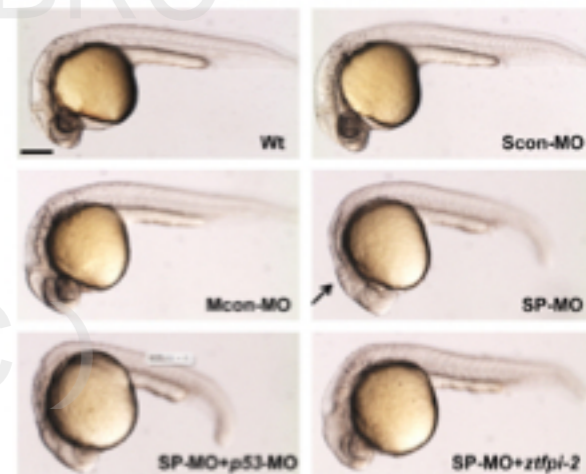
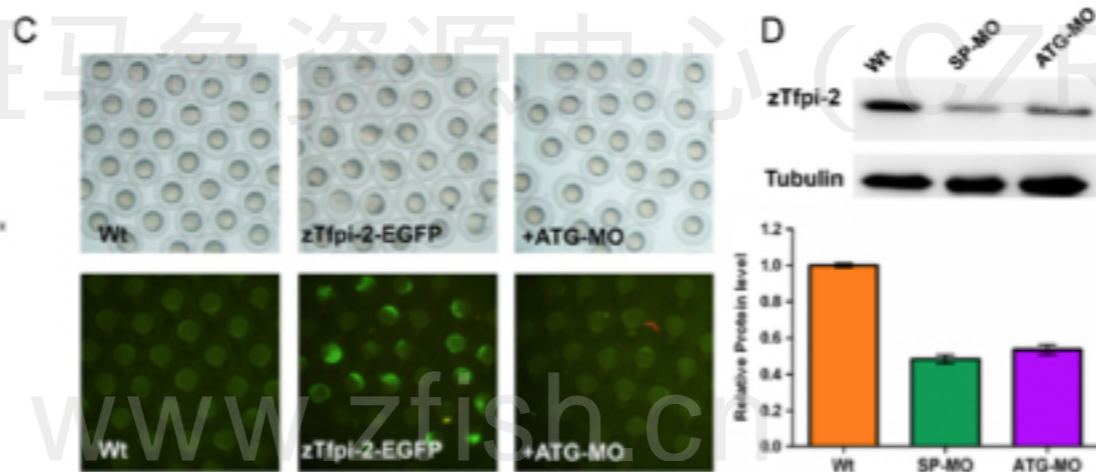
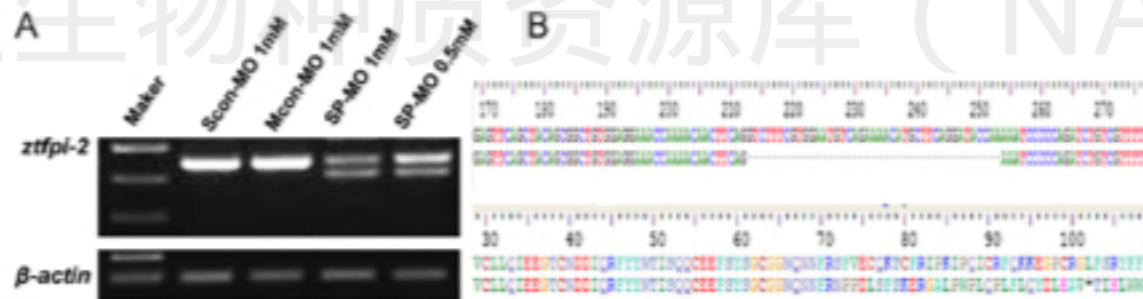


Morpholino工作原理 (Eisen JS, et al. Development. 2008.)

# 1、基因敲降技术

## Morpholino技术在斑马鱼研究中的应用

*ztfpi-2* (Tissue factor pathway inhibitor-2) gene  
 CNS development



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genetics 30:129-133 (2001)

Morpholino-Induced Knockdown of Zebrafish Engrailed Genes *eng2* and *eng3* Reveals Redundant and Unique Functions in Midbrain-Hindbrain Boundary Development

Steffen Scholpp and Michael Brand

Tissue factor pathway inhibitor-2: A novel gene involved in zebrafish central nervous system development

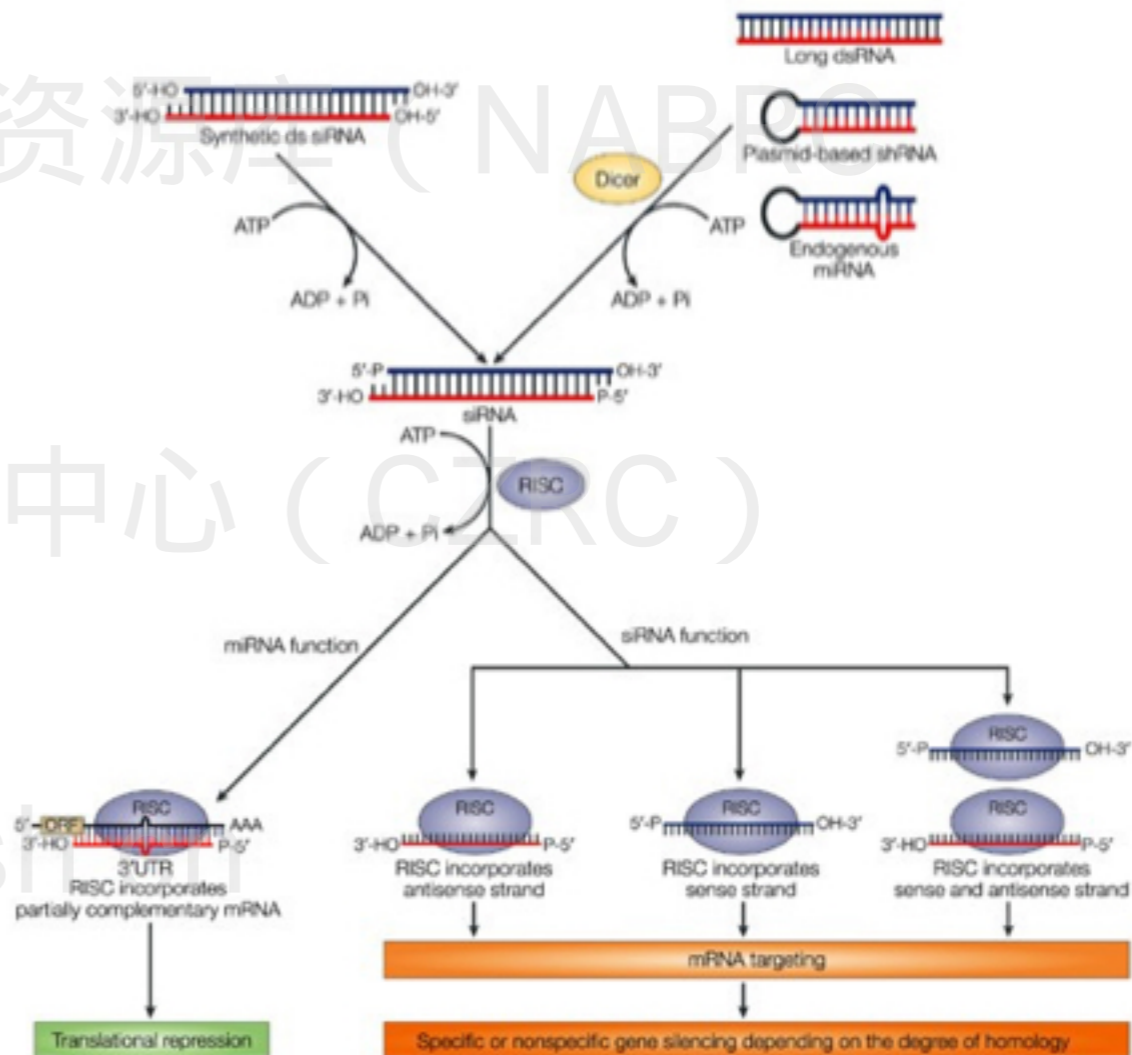
Yanli Zhang<sup>a</sup>, Lina Wang<sup>a</sup>, Wenhao Zhou<sup>b</sup>, Huijun Wang<sup>b</sup>, Jin Zhang<sup>a</sup>, Shanshan Deng<sup>a</sup>, Weihua Li<sup>c</sup>, Huawei Li<sup>a</sup>, Zuohua Mao<sup>a</sup>, Duan Ma<sup>a,b,\*</sup>

- *ztfpi-2* knockdown by MOs

# 1、基因敲降技术

## 1.2 RNAi技术:

- ① 人工合成的双链RNA序列 (dsRNA) 或siRNA 表达质粒体外合成;
- ② dsRNA经Dicer加工成21-25nt的siRNA;
- ③ siRNA中的反义链指导合成沉默复合体RISC;
- ④ RISC切割与siRNA反义链互补的mRNA分子, 引发其特异性降解, 从而阻断靶基因表达。



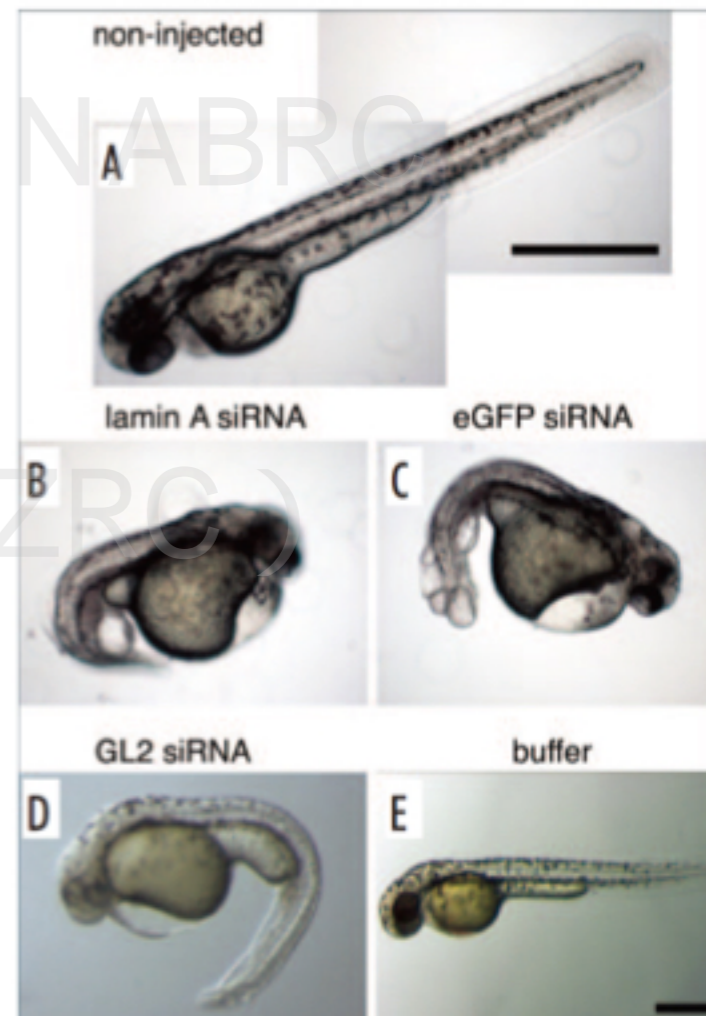
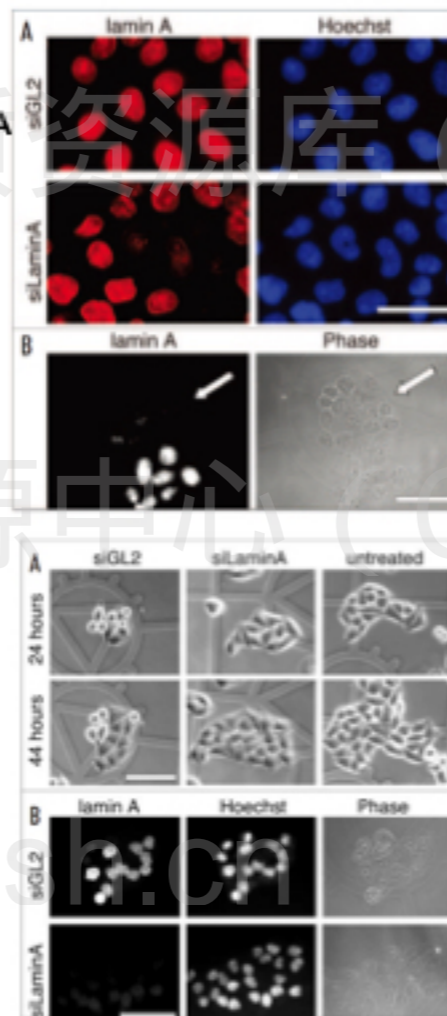
# 1、基因敲降技术

## RNAi技术在斑马鱼研究中的应用

- siRNAs work well in zebrafish cell lines (ZFL, SJD, ZF4)
- siRNA injected zebrafish embryos showed severe unspecific defects.

control firefly luciferase siRNA (siGL2)

ZFL cells



- siRNA injected embryos at 48 hpf showed severe unspecific defects.

RNA Biology

ISSN: 1547-6286 (Print) 1555-8584 (Online) Journal homepage: <https://www.tandfonline.com/loi/kmb20>

## Specific RNAi Mediated Gene Knockdown in Zebrafish Cell Lines

Jens Gruber, Heiko Manninga, Thomas Tuschl, Mary Osborn & Klaus Weber

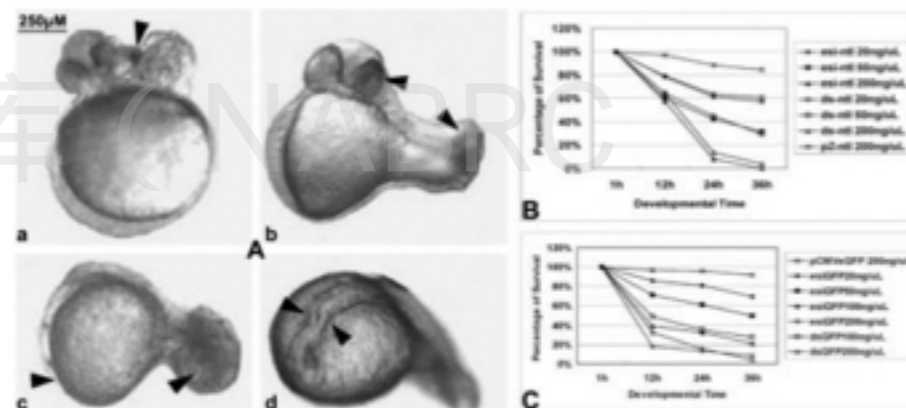
# 1、基因敲降技术

## RNAi技术在斑马鱼研究中的应用

*esi-ntl*: dsDNA of *ntl* digested by RNaseIII, purified

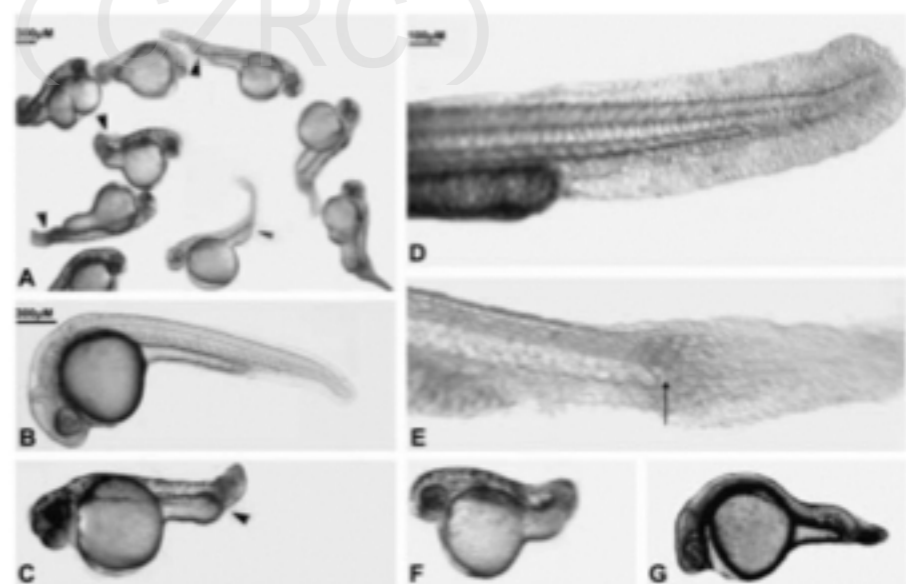
*si-ntl*: produced by SP6 RNA polymerase

*esi-ntl*, 50ng/ $\mu$ l



国家斑马鱼资源中心 (CZRC)

*si-ntl*, 4  $\mu$ g/ $\mu$ L



typical *ntl* mutant

Develop. Growth Differ. (2005) 47, 323–331

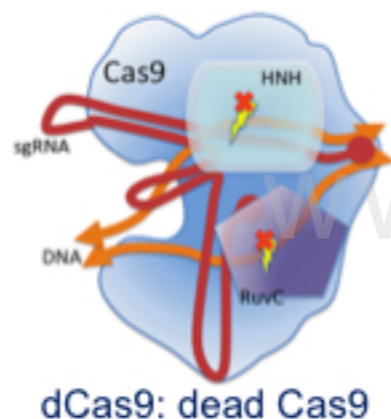
Efficient RNA interference in zebrafish embryos using siRNA synthesized with SP6 RNA polymerase

Wei-Yi Liu,<sup>1,2</sup> Yan Wang,<sup>1</sup> Yong-Hua Sun,<sup>1,\*</sup> Yun Wang,<sup>1,2</sup> Ya-Ping Wang,<sup>1</sup> Shang-Ping Chen<sup>1</sup> and Zuo-Yan Zhu<sup>1,\*</sup>

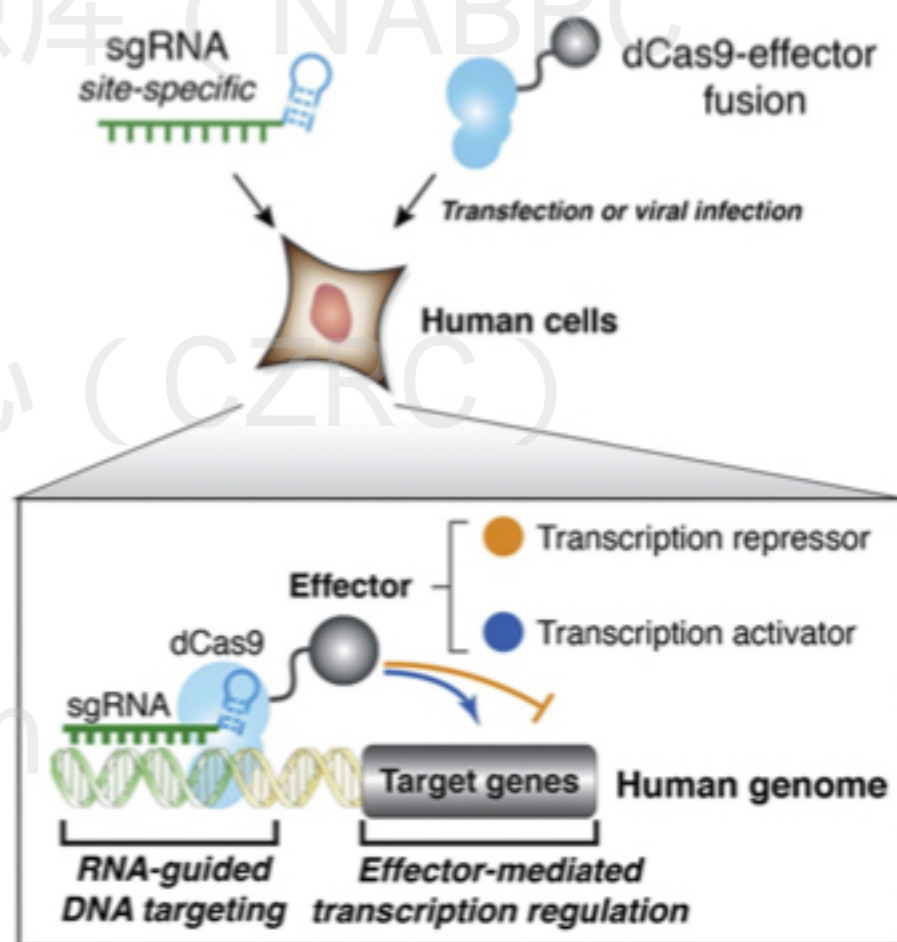
# 1、基因敲降技术

## 1.3 CRISPRi基因抑制技术:

- ① 失去切割DNA的功能的dCas9 (RuvC-D10A/HNH-H840A) 与转录抑制因子 (如Eve、KRAB) 形成融合蛋白;
- ② 根据目标基因的转录起始位点的特征, 设计 sgRNA 识别位点 (不少于 4 个);
- ③ CRISPRi系统可逆地抑制靶向基因转录的作用;
- ④ CRISPRi 敲降的有效性可以通过RT-PCR 法检测。



## CRISPRi: a modular RNA-guided genome regulation platform



(Gilbert LA, et al. Cell. 2013.)



# 1、基因敲降技术

	RNAi	反义Morpholino	CRISPRi
靶标的识别区域	21-23nt 的siRNA	25nt 的寡核苷酸单链	sgRNAs/dCas9蛋白
靶位点及作用	mRNA, 介导其降解	mRNA, 阻断其翻译或剪接	DNA转录起始位点, 抑制转录
作用对象	mRNA为主	mRNA	适用于多种转录本, 包括mRNA、非编码RNA、microRNA、反义转录本等
有效性评价	RT-PCR 法检测目标基因表达水平	RT-PCR 法检测目标基因表达水平或Western Blot检测目标蛋白表达水平	RT-PCR 法检测目标基因表达水平
优点	靶向精确, 操作方便	靶向精确、操作便捷	靶向精确、脱靶率低、细胞毒性低、价廉方便
缺点	脱靶率高、技术重现性差、具有细胞毒性	有一定的脱靶率, 存在诱发细胞凋亡风险、价格较高	作用受限于PAM序列

国家水生生物种质资源库 (NABRC)

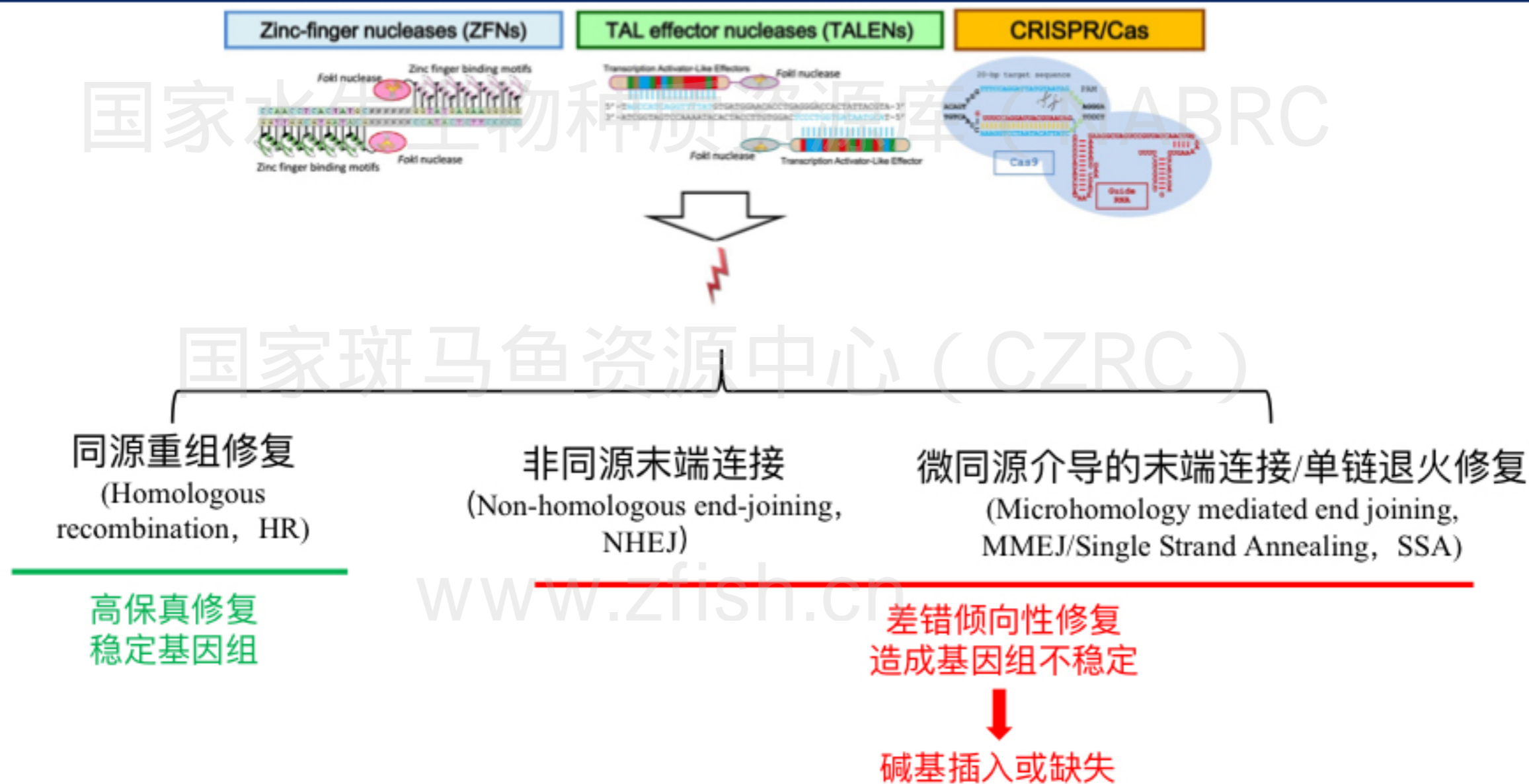
▶ 基因敲降技术 (Gene knockdown)

▶ DNA双链断裂修复 (DSB Repair) (CZRC)

▶ 基因编辑技术 (Gene Editing)

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## 2、DNA双链断裂修复

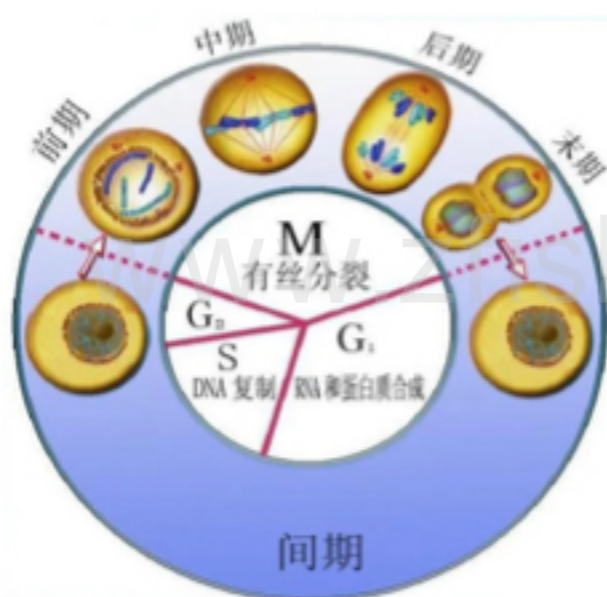


## 2、DNA双链断裂修复

### 2.1 同源重组修复

同源重组修复 (Homologous recombination, HR) 是保护基因组完整性的重要机制。以姐妹染色单体为DNA模板, 是无错误修复机制。只发生在S期和G<sub>2</sub>期。

- ① DNA双链断裂处的识别和处理;
- ② RAD51-ssDNA的侵入;
- ③ DNA的修复。

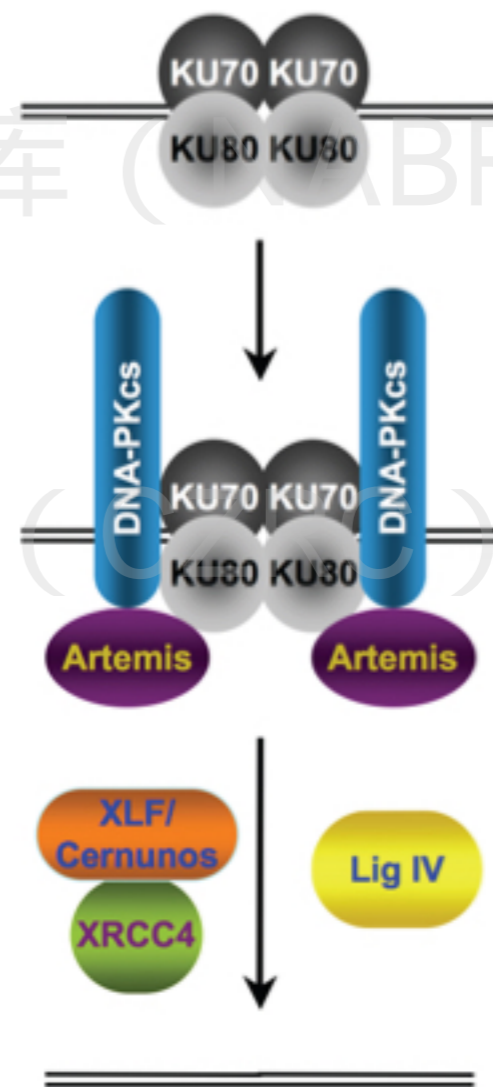


## 2、DNA双链断裂修复

### 2.2 非同源末端连接

非同源末端连接 (Non-homologous end-joining, NHEJ) 依赖于Ku70-Ku80和DNA连接酶 IV-XRCC4异源二聚体来修复DSB, 在连接处随机引入小的突变序列。

- ① DNA双链断裂处的识别和结合;
- ② 断裂末端的加工处理;
- ③ DNA末端与连接, 完成修复过程。



NHEJ

(Anastazja G., et al., *Am J Cancer Res* 2012)

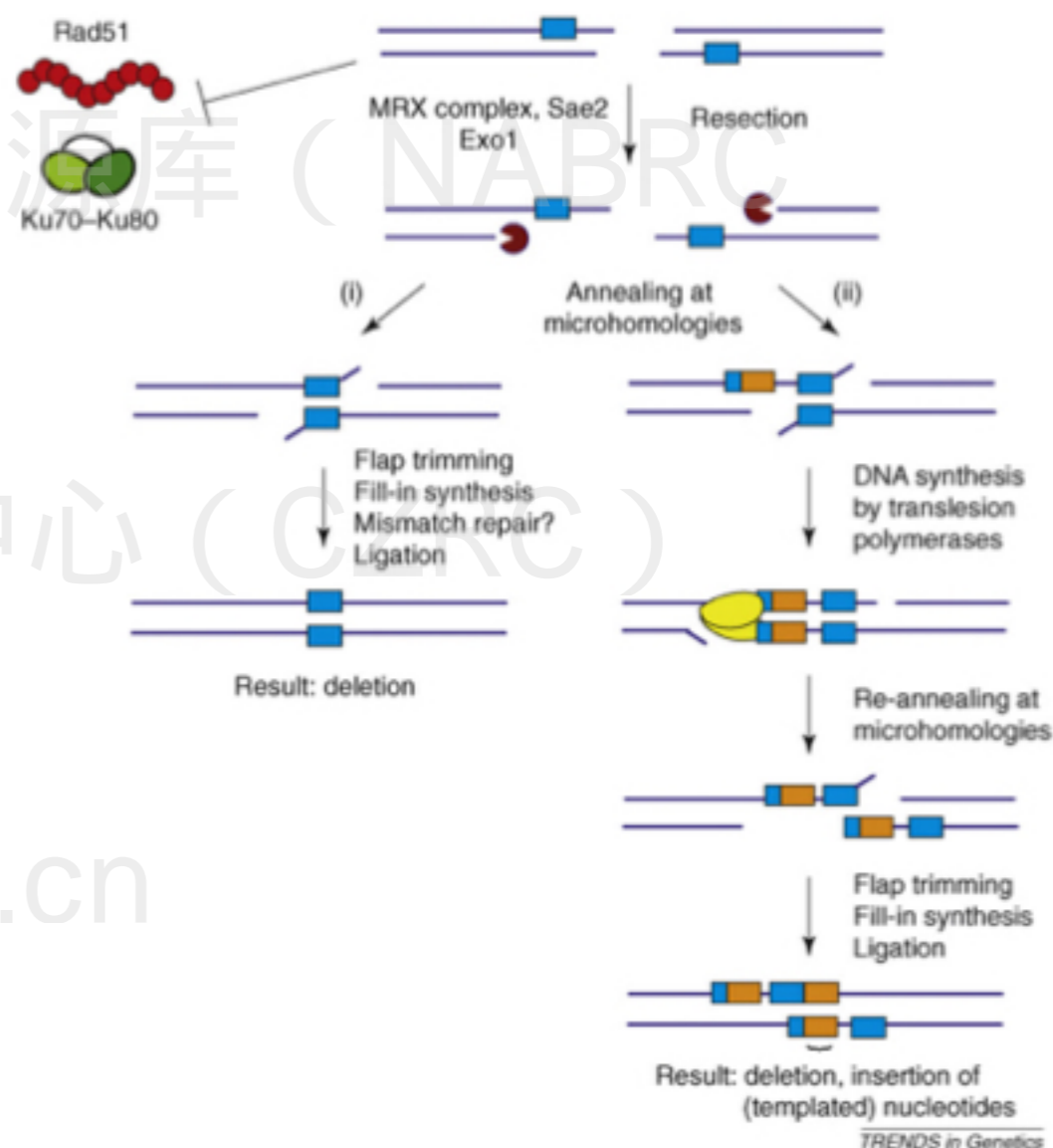
## 2、DNA双链断裂修复

### 2.3 微同源介导的末端连接/单链退火修复

微同源介导的末端连接/单链退火修复

(Microhomology mediated end joining, MMEJ / Single Strand Annealing, SSA) 利用同向的同源序列 (5–25 bp/MMEJ; >30bp/SSA) 来介导末端连接的修复机制，差错倾向性修复过程。MMEJ修复是在G1 /早S期活跃。

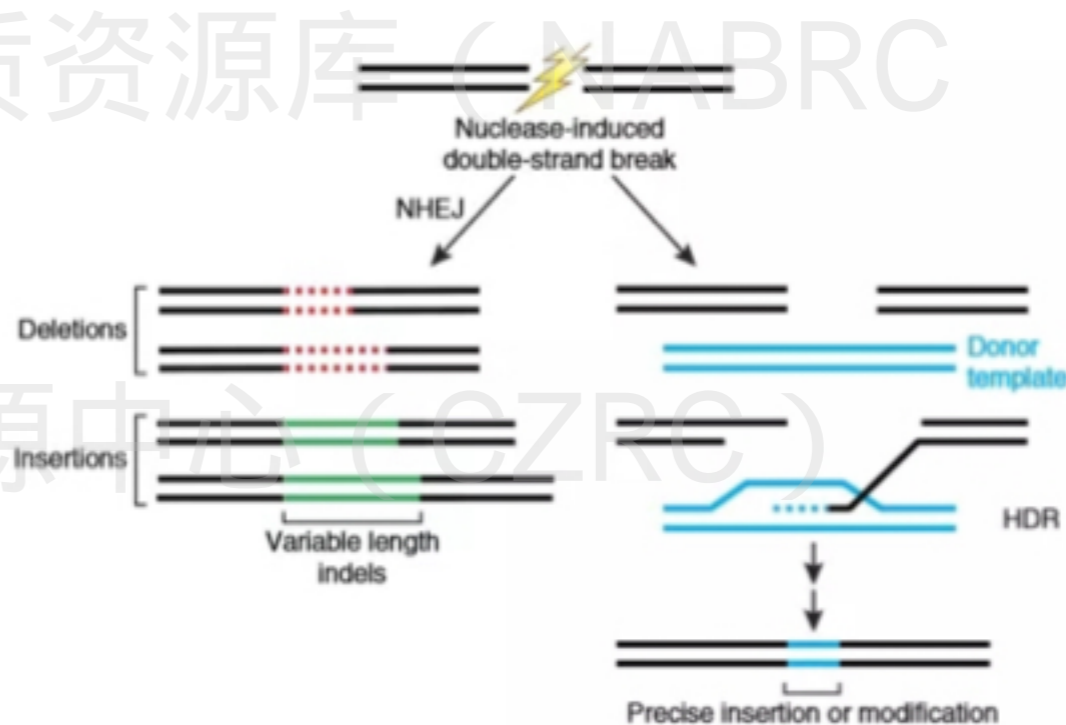
- ① DNA双链断裂末端识别；
- ② 5'末端处理；
- ③ 微同源序列相互退火，完成修复过程。



## 2、DNA双链断裂修复

### 2.4 同源定向修复

同源定向修复 (Homology directed repair, HDR) 是细胞内一种修复DNA双链损伤的机制, 当细胞核内存在与损伤DNA同源的DNA片段时, 机体将以该同源DNA片段作为模板对DNA损伤进行修复。



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▶ 基因敲降技术 (Gene knockdown)

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### 3、基因编辑技术

基因突变技术是人为地使靶基因的序列发生碱基对的增加、缺失或替换，引起的靶基因结构的改变，以定点修饰改造特定基因，研究基因功能。斑马鱼突变品系广泛应用于遗传学、发育生物学、细胞生物学、医学、环境毒理学、水产育种学等研究领域。



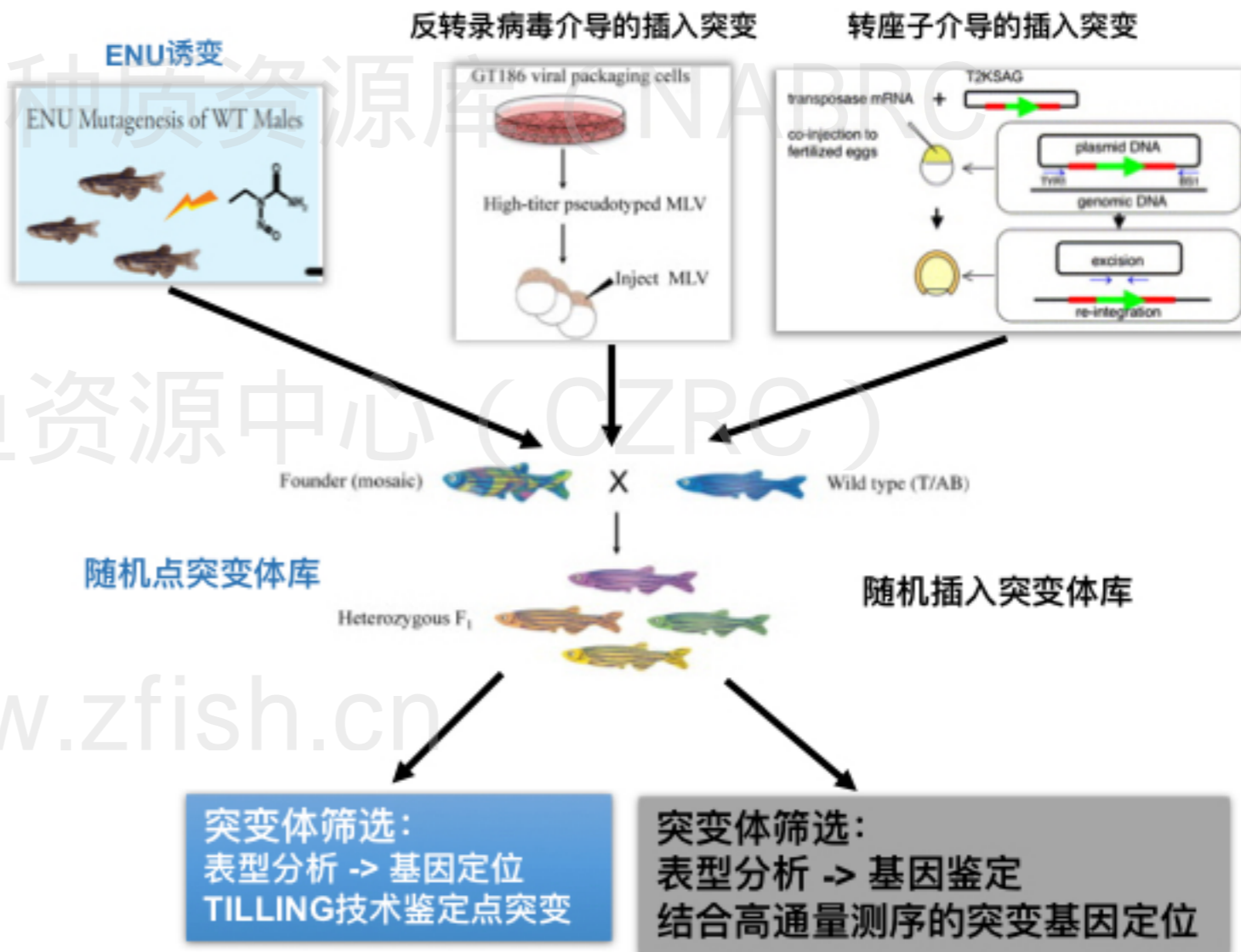
[www.zfish.cn](http://www.zfish.cn)

# 3、基因编辑技术

## 正向遗传学技术：

正向遗传学技术是指通过生物个体的基因组的自发突变或人工诱变，寻找相关的表型或性状的变化，然后从这些特定性状变化的个体中找到对应的突变基因并揭示其功能。

- ① 突变位点随机；
- ② 获得大量早期发育突变品系；
- ③ 突变体筛选繁琐耗时。



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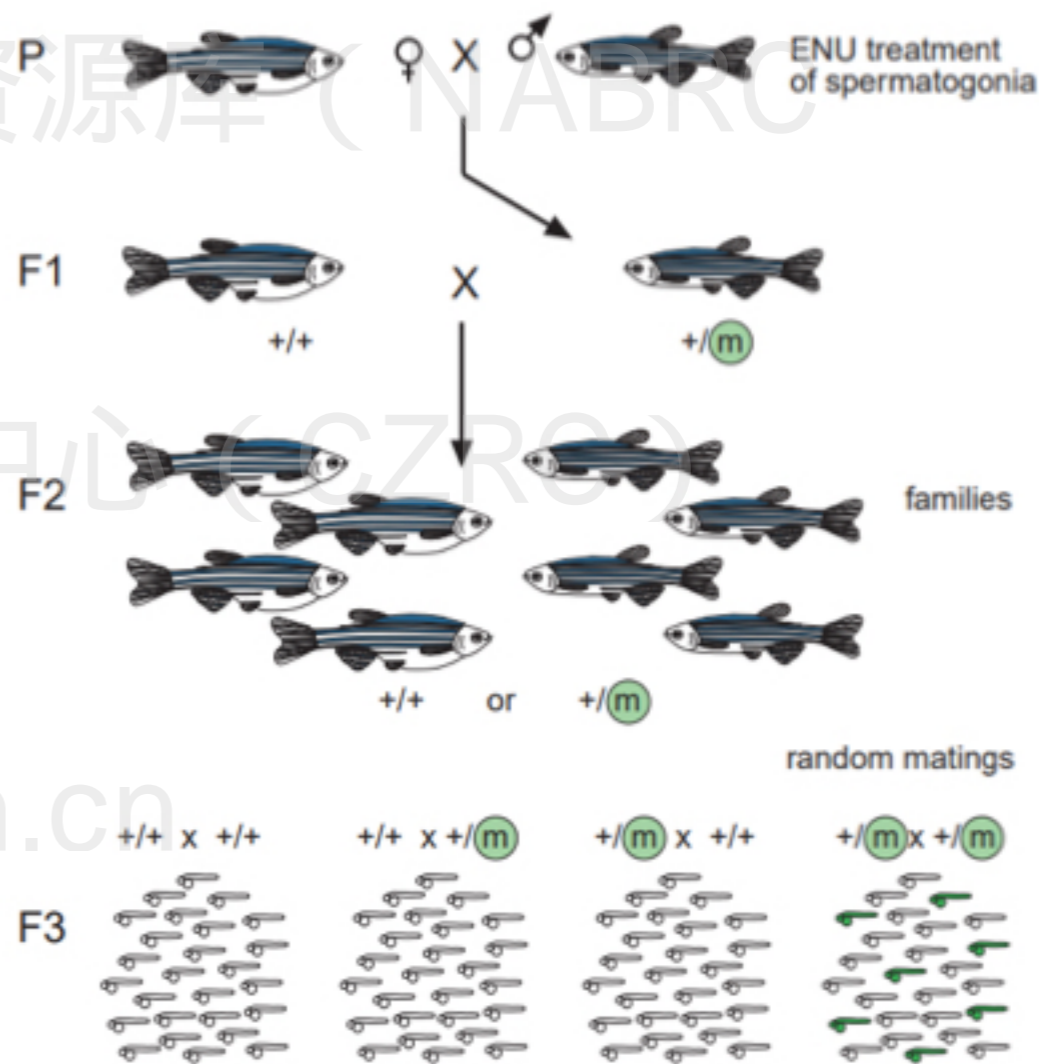
# 3、基因编辑技术

## 斑马鱼大规模ENU诱变

Development 123, 1-36  
Printed in Great Britain © The Company of Biologists Limited 1996  
DEV3343

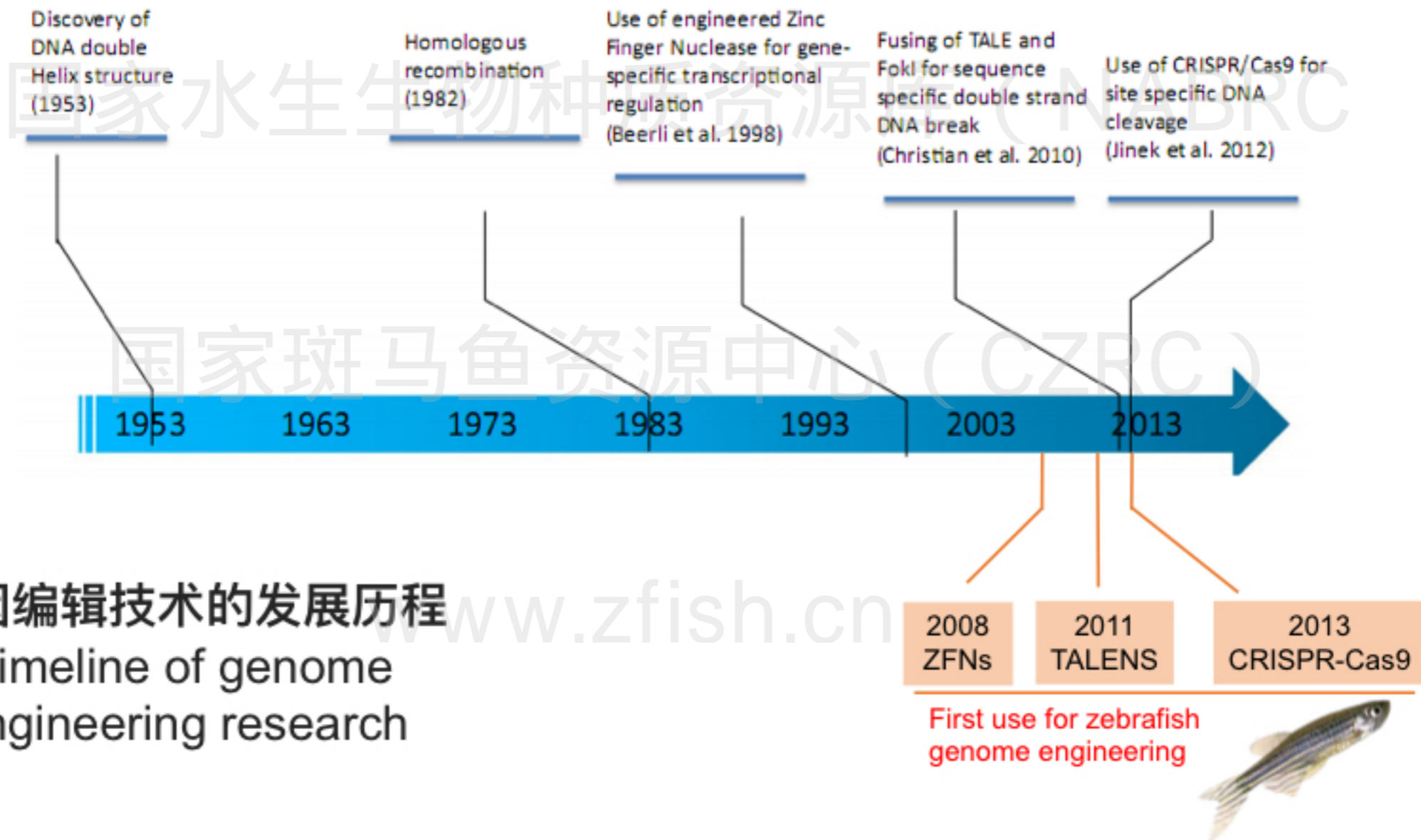
The identification of genes with unique and essential functions in the development of the zebrafish, *Danio rerio*

Pascal Haffter, Michael Granato<sup>2</sup>, Michael Brand<sup>1</sup>, Mary C. Mullins<sup>2</sup>, Matthias Hammerschmidt<sup>3</sup>, Donald A. Kane<sup>1</sup>, Jörg Odenthal, Fredericus J. M. van Eeden, Yun-Jin Jiang, Carl-Philipp Heisenberg, Robert N. Kelsh<sup>1</sup>, Makoto Furutani-Seiki, Elisabeth Vogelsang<sup>2\*</sup>, Dirk Beuchle<sup>1†</sup>, Ursula Schach, Cosima Fabian and Christiane Nüsslein-Volhard\*



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# 3、基因编辑技术



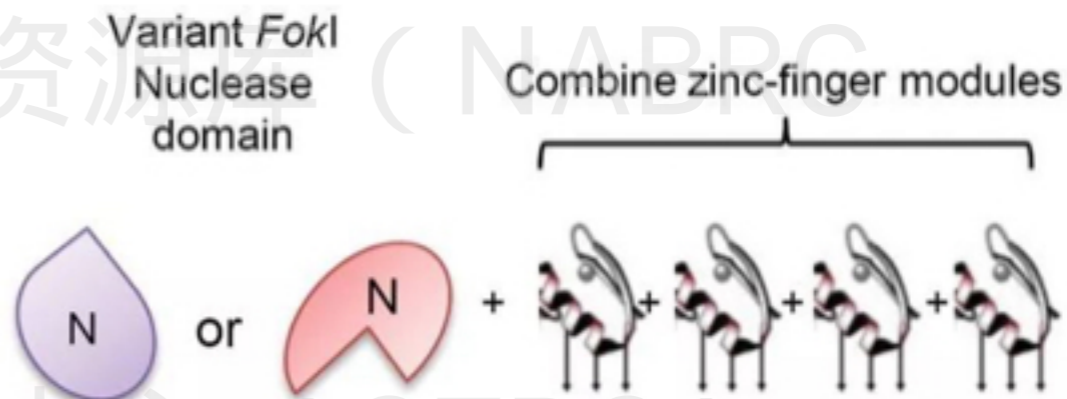
- 基因编辑技术的发展历程  
Timeline of genome engineering research

# 3、基因编辑技术

## 3.1 ZFNs技术

① ZFN=锌指DNA结合域 ZF+核酸内切酶 Fok I;

锌指核酸酶 (Zinc-finger nuclease, **ZFN**) 是一类人工合成的限制性内切酶，由锌指DNA结合域 (zinc finger DNA-binding domain) 与核酸内切酶的DNA切割域 (DNA cleavage domain) 融合而成。

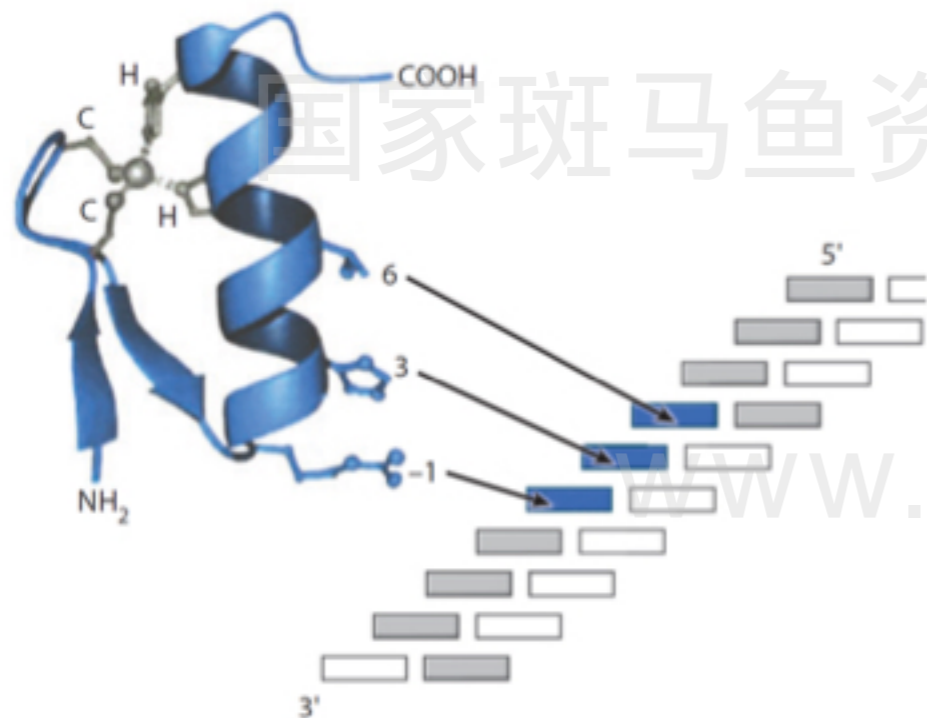


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# 3、基因编辑技术

## 3.1 ZFNs技术

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- ② ZF: DNA 识别, 3-4个 Cys2-His2锌指蛋白串联组成, 每个锌指蛋白识别一个特异的三联体碱基;



Variant FokI  
Nuclease  
domain

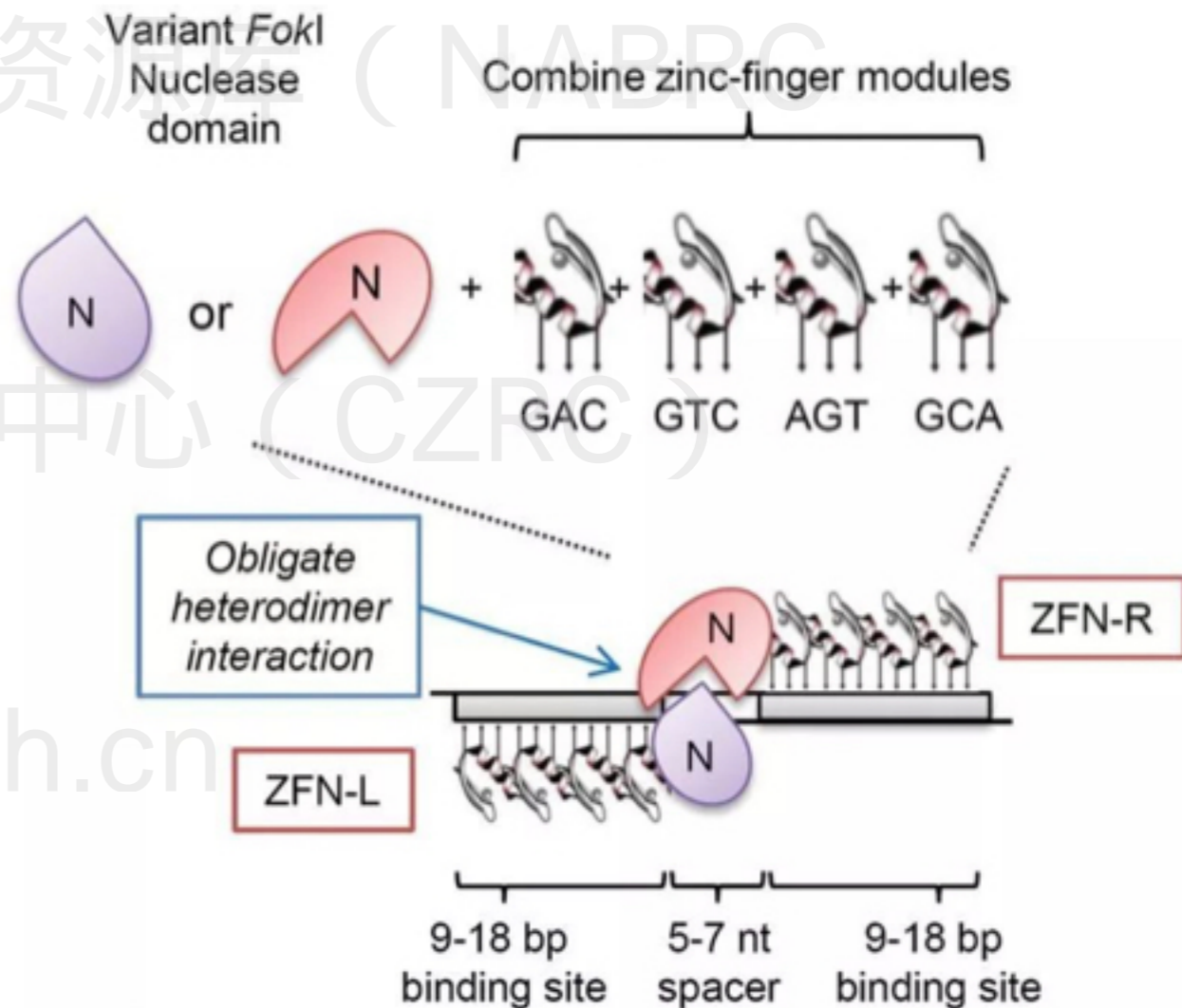


锌指蛋白负责DNA识别

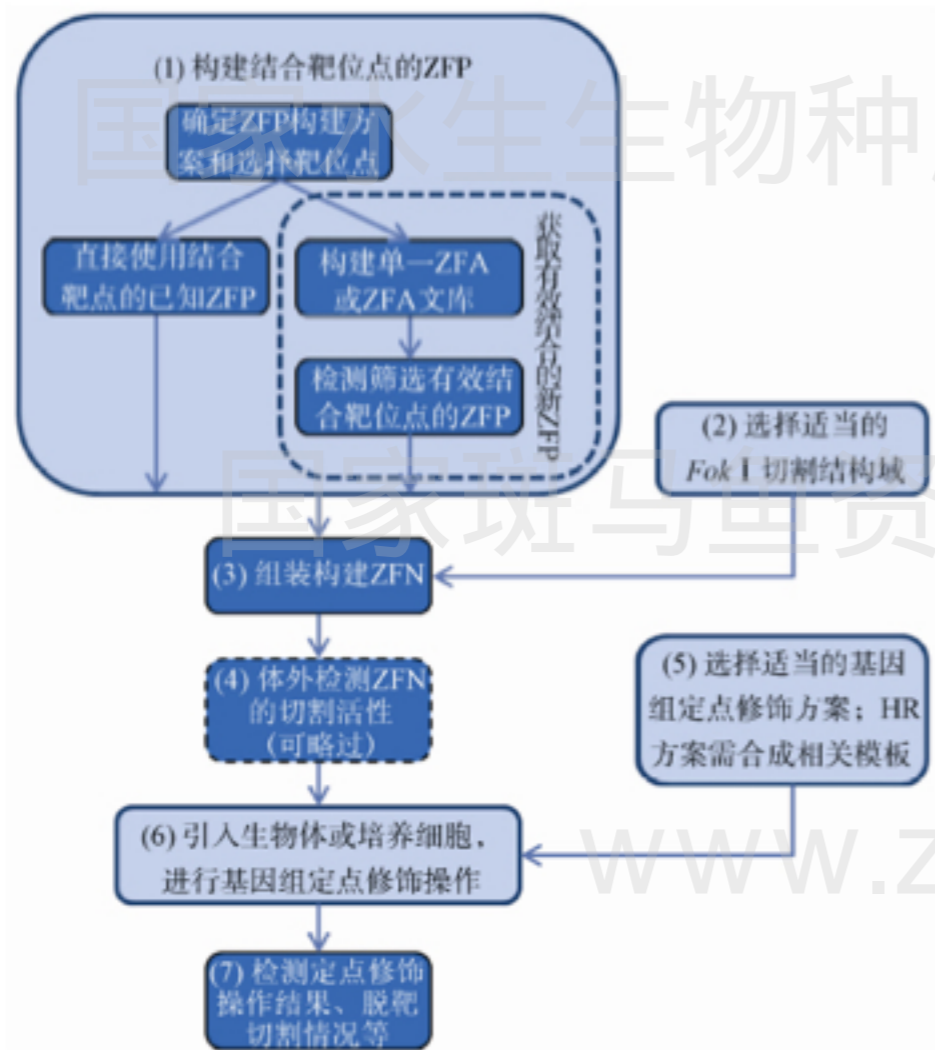
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- ③ Fok I: 形成二聚体对DNA定向剪切。  
间隔区spacer: 以5~7bp为宜



### 3、基因编辑技术



#### ZFN设计相关网站

ZifBase, <http://web.iitd.ac.in/~sundar/zifbase>

ZiFiT, <http://zifit.partners.org/ZiFiT>

<http://mccb.umassmed.edu/ZFPsearch.html>

<http://www.addgene.org/zfc>

ZiFDB, <http://bindr.gdcb.iastate.edu:8080/ZiFDB>

- ZFNs实验构建流程 (肖安等, 遗传, 2011)



# 3、基因编辑技术

## ZFN技术在斑马鱼研究中的应用

- 2008年首次成功地使用ZFN基因编辑技术，对斑马鱼基因组进行精确的位点特异性编辑。

*Nat Biotechnol.* 2008 June ; 26(6): 702-708. doi:10.1038/nbt1409.

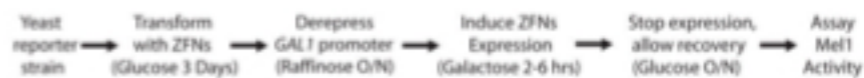
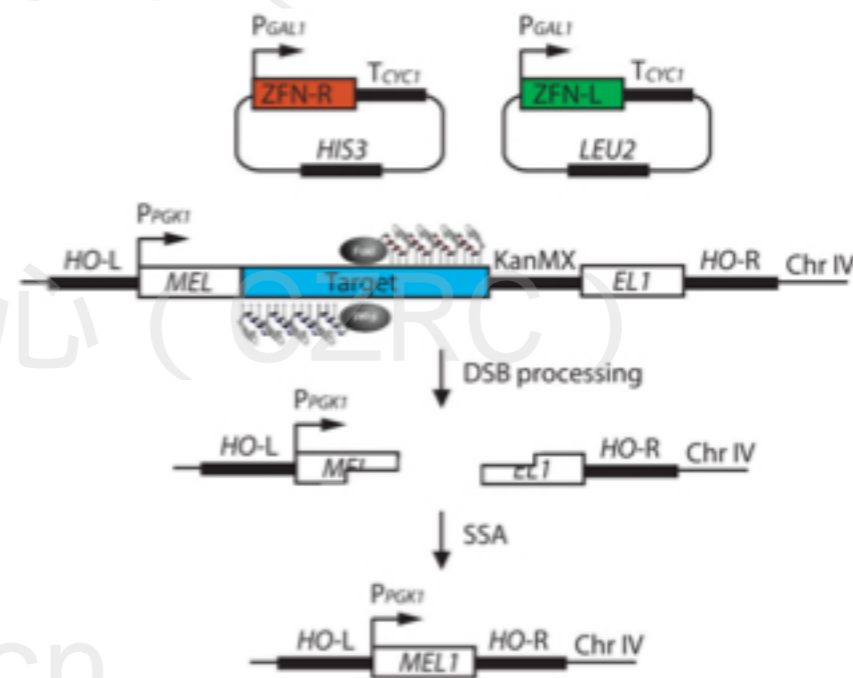
### Heritable Targeted Gene Disruption in Zebrafish Using Designed Zinc Finger Nucleases

Yannick Doyon<sup>2,4</sup>, Jasmine M McCammon<sup>1,4</sup>, Jeffrey C Miller<sup>2</sup>, Farhoud Faraji<sup>2</sup>, Catherine Ngo<sup>2</sup>, George E Katibah<sup>2</sup>, Rainier Amora<sup>2</sup>, Toby D Hocking<sup>2</sup>, Lei Zhang<sup>2</sup>, Edward J Rebar<sup>2</sup>, Philip D Gregory<sup>2</sup>, Fyodor D Urnov<sup>1,2</sup>, and Sharon L Amacher<sup>1,3</sup>

*Nat Biotechnol.* 2008 June ; 26(6): 695-701.

### Targeted gene inactivation in zebrafish using engineered zinc finger nucleases

Xiangdong Meng<sup>1,4</sup>, Marcus B. Noyes<sup>1,2</sup>, Lihua (Julie) Zhu<sup>1</sup>, Nathan D. Lawson<sup>1,3</sup>, and Scot A. Wolfe<sup>1,2</sup>



- Yeast-based system of identification of maximally active ZFNs

# 3、基因编辑技术

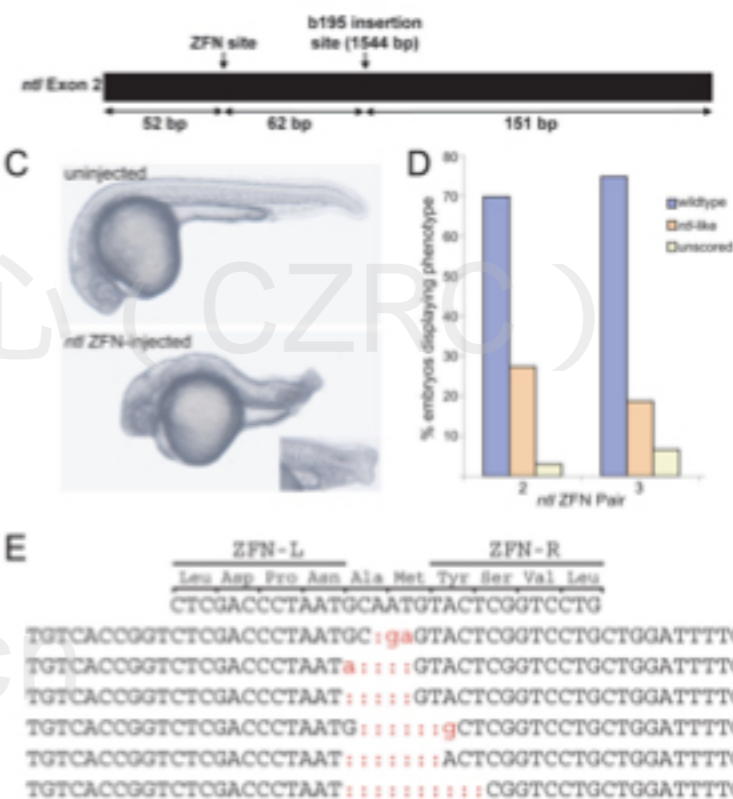
## ZFN技术在斑马鱼研究中的应用

- 2008年首次成功地使用ZFN基因编辑技术，对斑马鱼基因组进行精确的位点特异性编辑。

*Nat Biotechnol.* 2008 June ; 26(6): 702-708. doi:10.1038/nbt1409.

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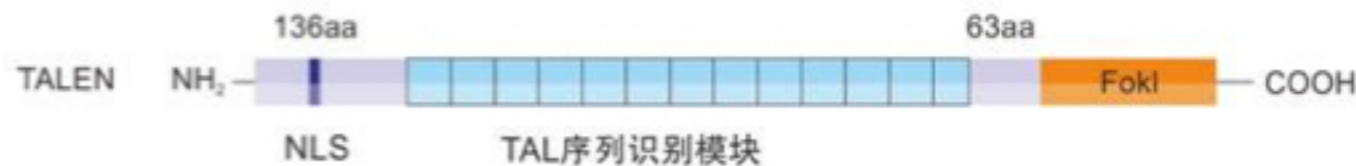


- Injection of no tail ZFN-encoding mRNA into zebrafish embryos induces targeted loss-of-function mutations in somatic cells

# 3、基因编辑技术

## 3.2 TALENs技术

① TALEN=TALE+核酸内切酶Fok I;



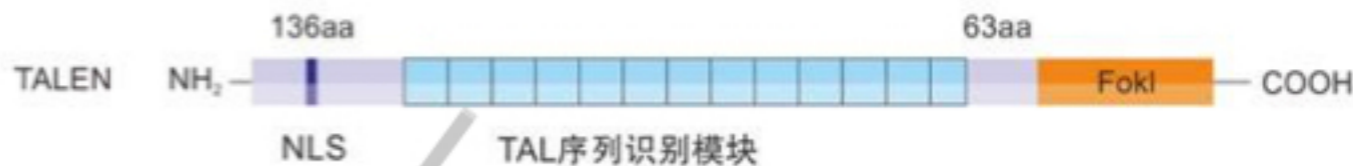
**TALEN** (transcription activator-like effector nuclease) 是一类人工合成的限制性内切酶，由可编码的串联TALE蛋白域与核酸内切酶的DNA切割域 (DNA cleavage domain) 融合而成。

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# 3、基因编辑技术

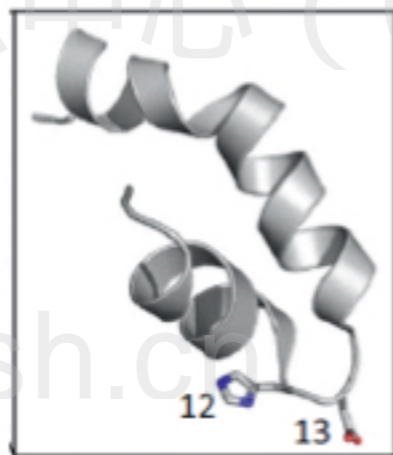
## 3.2 TALENs技术

- ① TALEN=TALE+核酸内切酶Fok I;
- ② TALE: DNA 识别, 一串TALE蛋白组成, TALE蛋白识别特异碱基对;



**TALE** (transcription activator-like effector)

AvrBs3蛋白 (*Xanthomonas*, 黄单胞菌), 含17.5个由34个氨基酸组成的重复模块



TAL蛋白

氨基酸残基与碱基的对应关系:

**NG** -> T

**HD** -> C

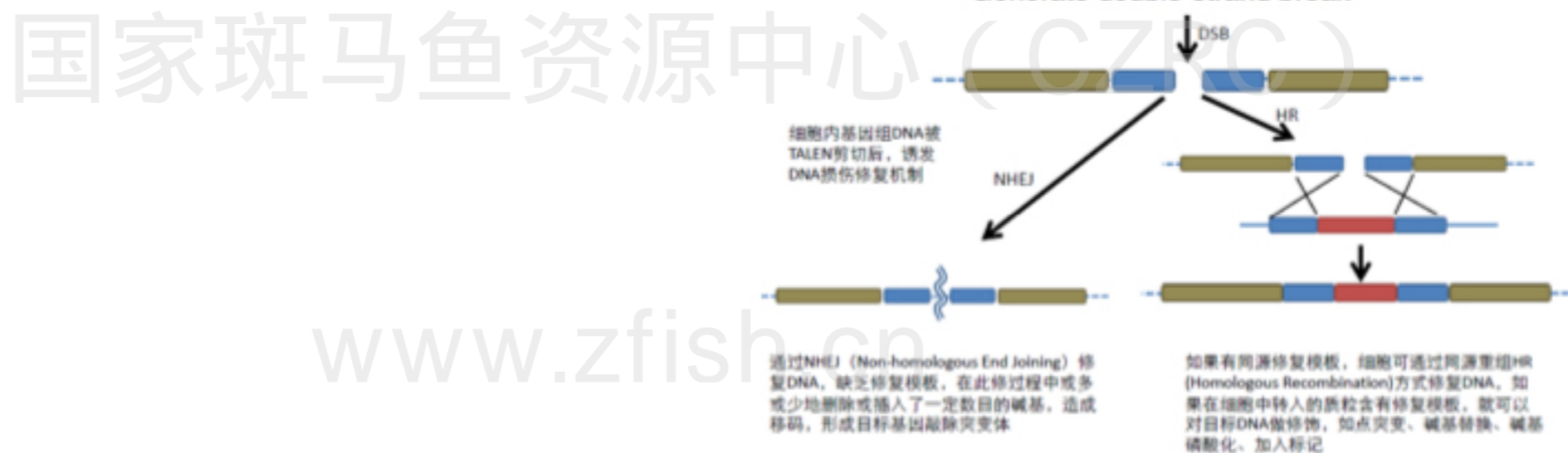
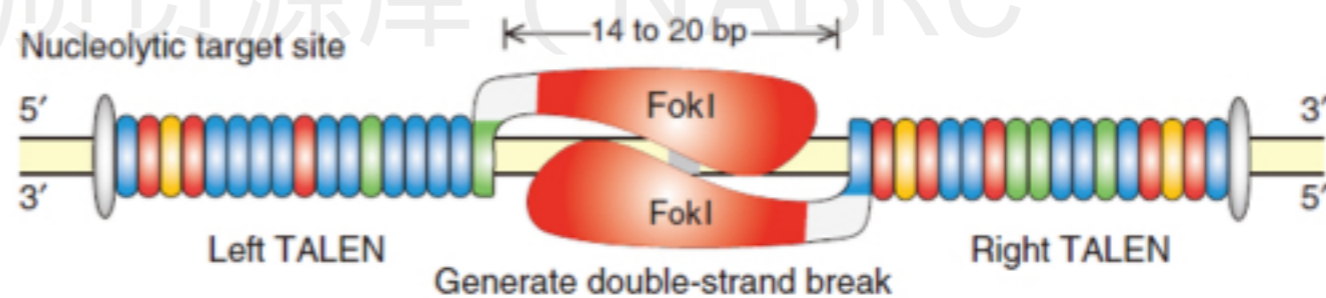
**NI** -> A

**NN** -> G (> (NK->G))

# 3、基因编辑技术

## 3.2 TALENs技术

- ① TALEN=TALE+核酸内切酶Fok I;
- ② TALE: DNA 识别, 一串TALE蛋白组成, TALE蛋白识别特异碱基对;
- ③ Fok I: 形成二聚体对DNA定向剪切。



# 3、基因编辑技术

## 3.2 TALENs技术

- ① TALEN=TALE+核酸内切酶Fok I;
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- ③ Fok I: 形成二聚体对DNA定向剪切。

氨基酸残基与碱基的对应关系:

**NG** -> **T**

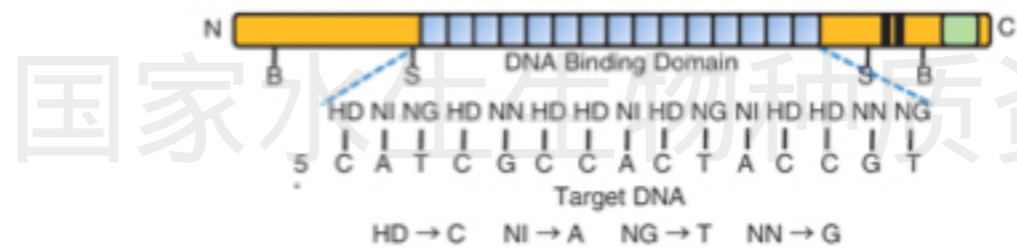
**HD** -> **C**

**NI** -> **A**

**NN (>NK)** -> **G**

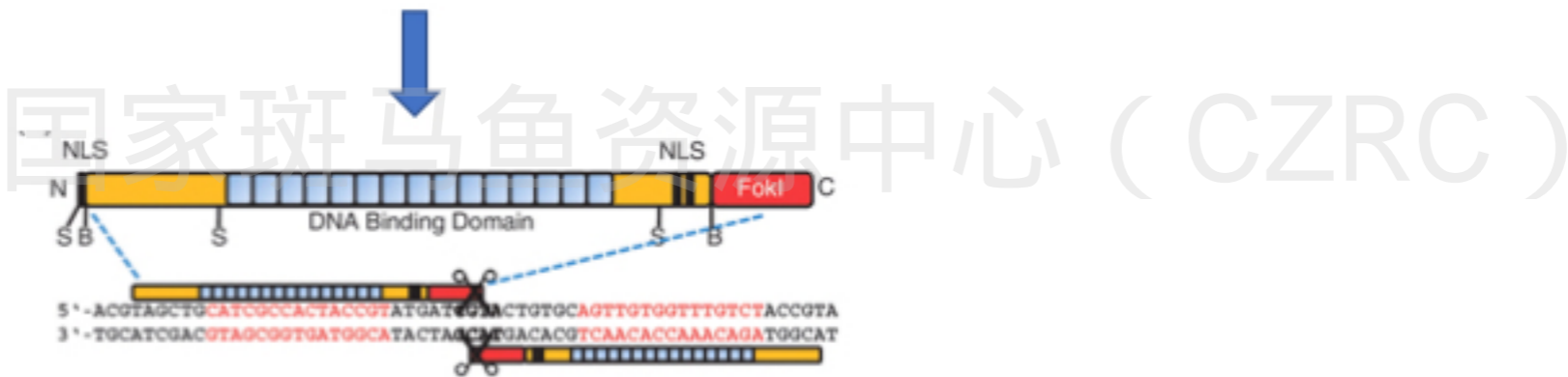


# 3、基因编辑技术



## • 基于TALENs技术的基因敲除品系构建流程

- TALENs靶点识别模块构建



- TALENs表达质粒的构建

合成TALEN mRNA，注射到斑马鱼胚胎中，获取阳性F0代

突变体筛选：历经2代，酶切检测法/PCR测序

# 3、基因编辑技术

## TALENs技术在斑马鱼中的应用

- 2011年，首次成功地使用TALEN基因编辑技术，对斑马鱼基因组进行精确的基因编辑。

### Heritable gene targeting in zebrafish using customized TALENs

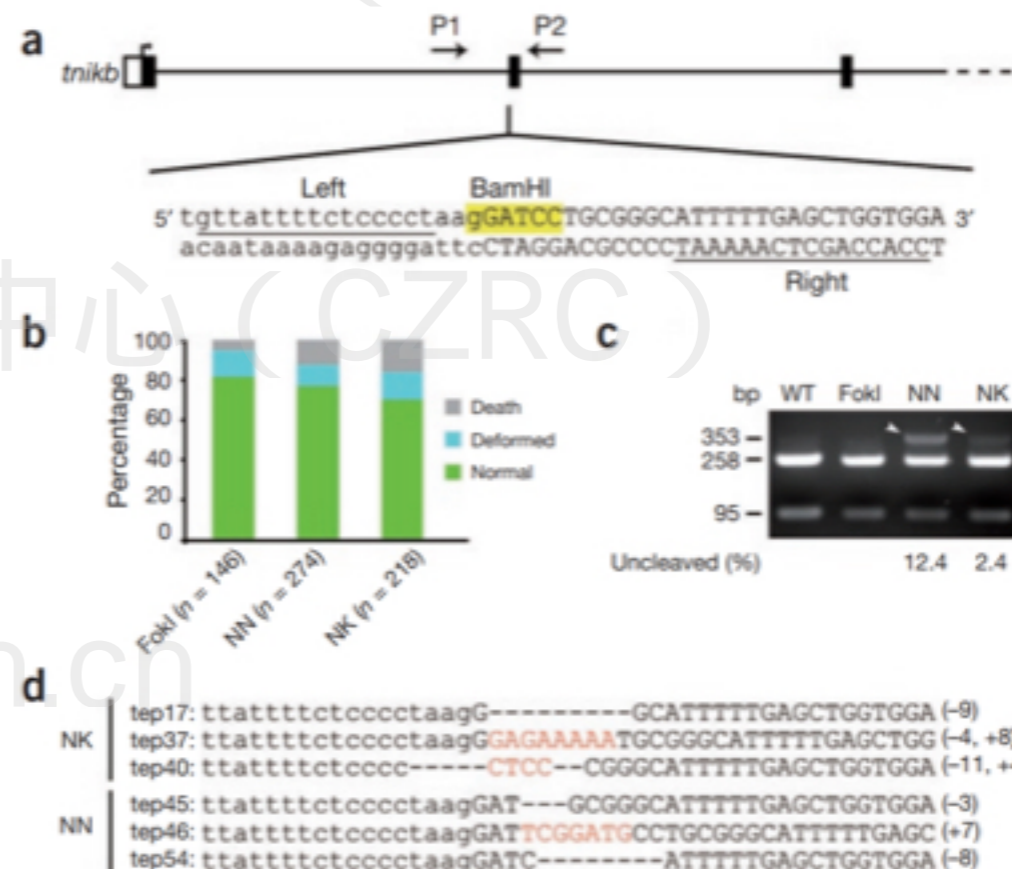
Peng Huang, An Xiao, Mingguo Zhou, Zuoyan Zhu, Shuo Lin & Bo Zhang

Nature Biotechnology 29, 699–700 (2011) | Cite this article

Nat Biotechnol. ; 29(8): 697–698. doi:10.1038/nbt.1934.

### Targeted gene disruption in somatic zebrafish cells using engineered TALENs

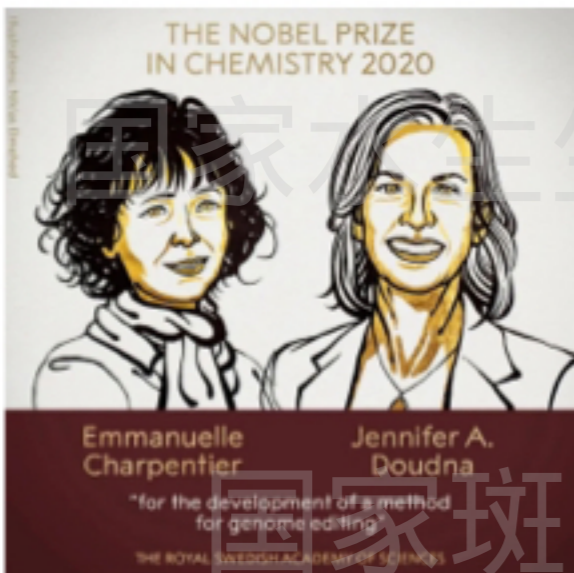
Jeffrey D. Sander<sup>1,2,\*</sup>, Lindsay Cade<sup>3,\*</sup>, Cyd Khayter<sup>1,2</sup>, Deepak Reyon<sup>4</sup>, Randall T. Peterson<sup>3,5</sup>, J. Keith Joung<sup>1,2,#</sup>, and Jing-Ruey J. Yeh<sup>3,#</sup>



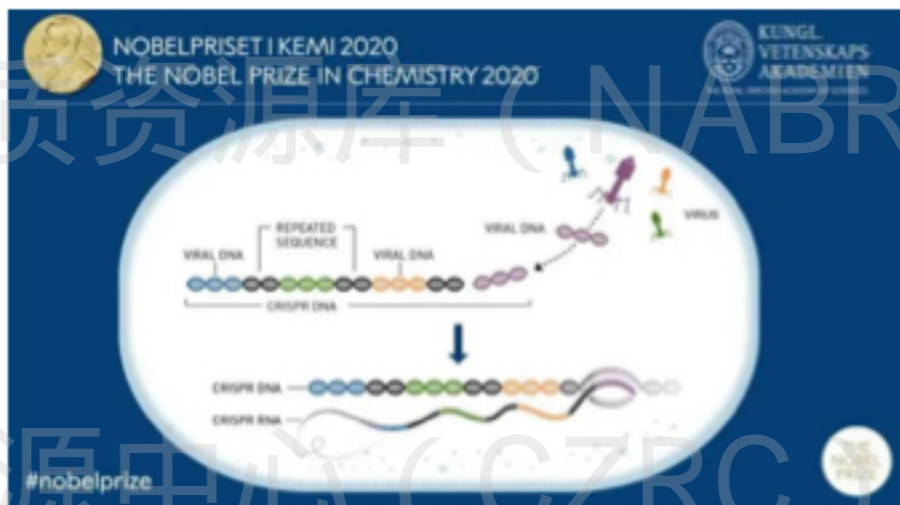
- Targeted and heritable disruption of the *tnika* gene in zebrafish using TALENs. (Huang P et al, Nat Biotechnol. 2011)



# 3、基因编辑技术



2020年诺贝尔化学奖



CRISPR/Cas 系统

1987年，第一次发现  
CRISPR

2005-2006年，证实  
CRISPR中含有病毒序  
列

2008-2010年，证明二型  
CRISPR-Cas切割DNA

2012年，CRISPR/Cas9  
编辑技术诞生

2002年，统一命  
名为CRISPR-  
Cas

2007年，证明CRISPR-  
Cas为细菌适应性免疫系  
统

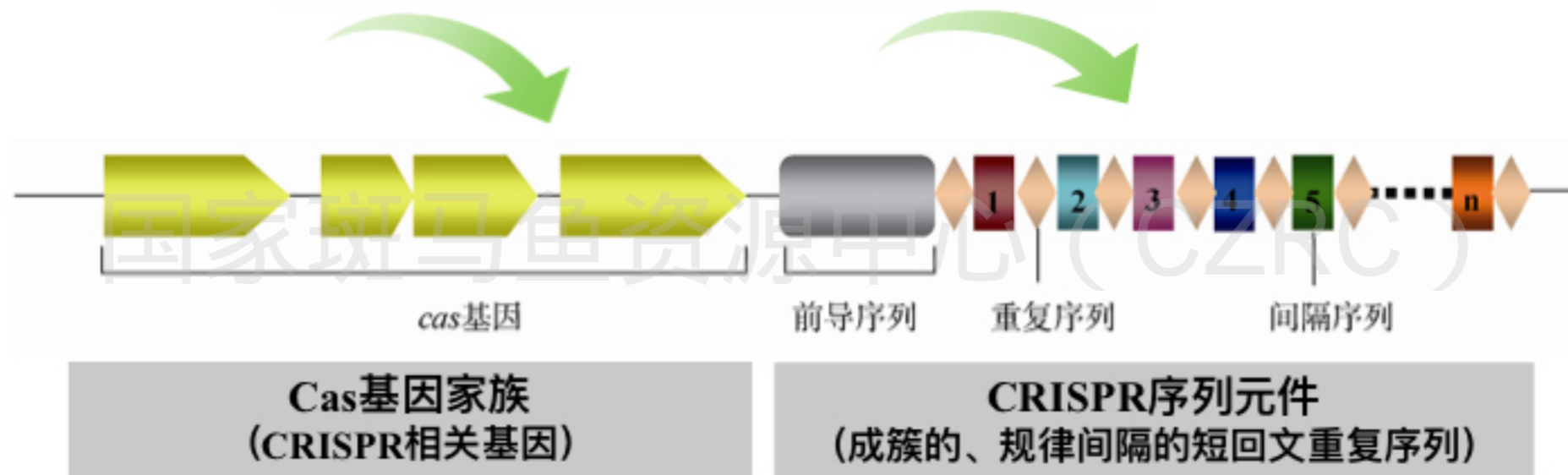
2011年，发现tracr RNA, cas9是  
二型CRISPR-Cas所需唯一基因

### 3、基因编辑技术

#### 细菌获得性免疫：CRISPR-Cas system

国家水生生物种质资源库 (NABRC)

CRISPR-Cas主要由两部分组成：

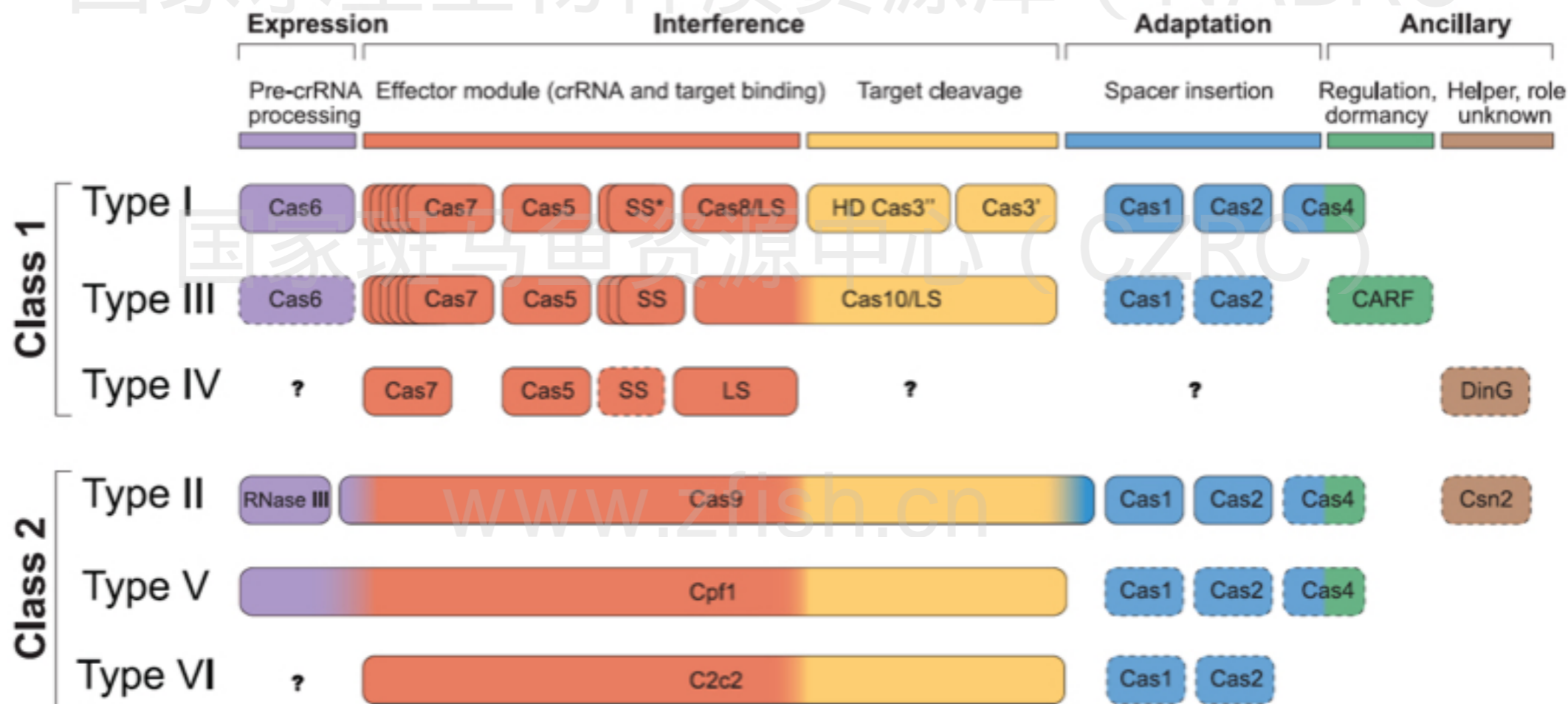


**CRISPR** (clustered, regularly interspaced, short palindromic repeats, 成簇的规律间隔的短回文重复序列):

**Cas基因** (CRISPR-associated protein): 位于CRISPR位点附近, 是一个较大的多态性家族, 编码的蛋白具有核酸酶相关的功能域。

# 3、基因编辑技术

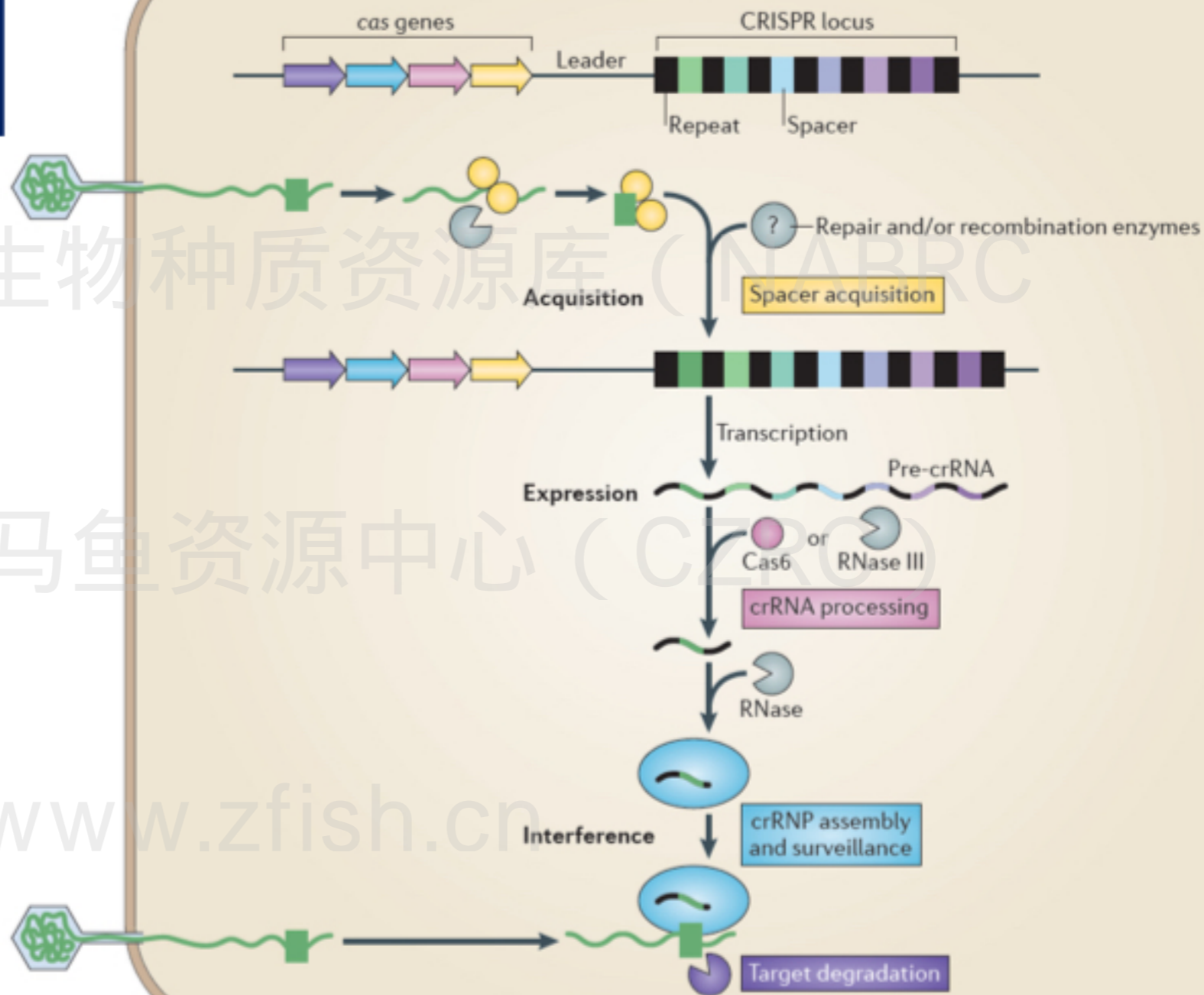
## 细菌获得性免疫: CRISPR-Cas system



# 3、基因编辑技术

## CRISPR-Cas system作用机制

1. Spacer的获得
2. 合成CrRNA
3. DNA interference



( John O.. Curr Opin Struct Biol. 2014)

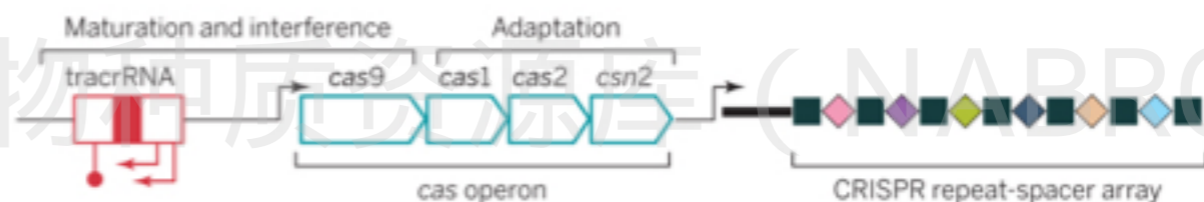
# 3、基因编辑技术

## CRISPR-Cas9 system 工作原理

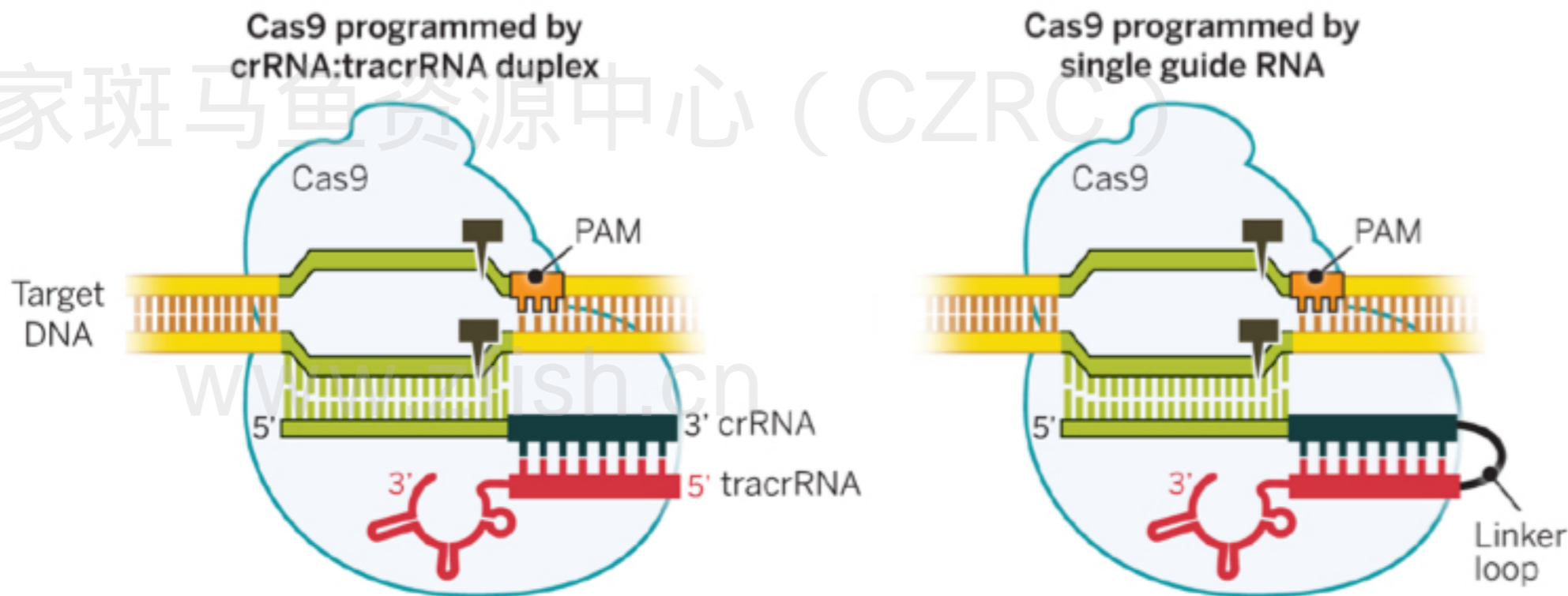
### CRISPR/Cas9技术:

利用细菌获得性免疫CRISPR-cas系统进行基因组序列操作，gRNA与cas9蛋白（具有HNH和RuvC两个具核酸内切酶活性区域）形成复合体后引导cas9蛋白对与gRNA互补的一段基因组序列进行剪切，结合机体修复系统对基因组进行编辑。

A Genomic CRISPR locus



B tracrRNA:crRNA co-maturation and Cas9 co-complex formation



(Jennifer A. D. science. 2016)

### 3、基因编辑技术

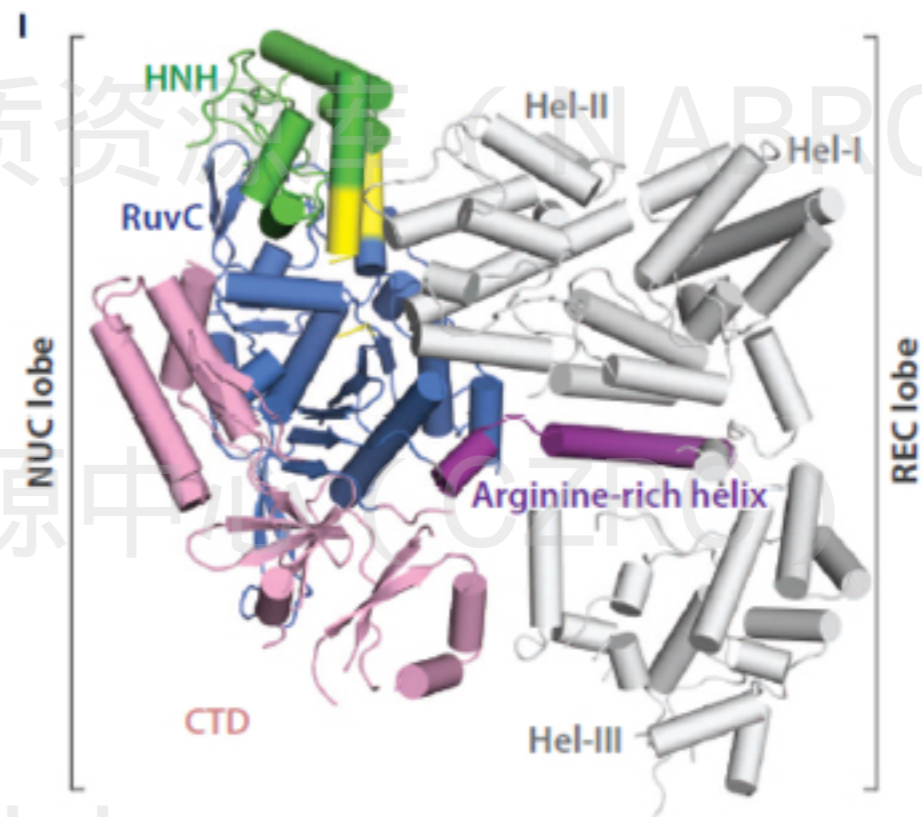
#### SpCas9蛋白结构

(*Streptococcus pyogenes*, 酿脓链球菌)

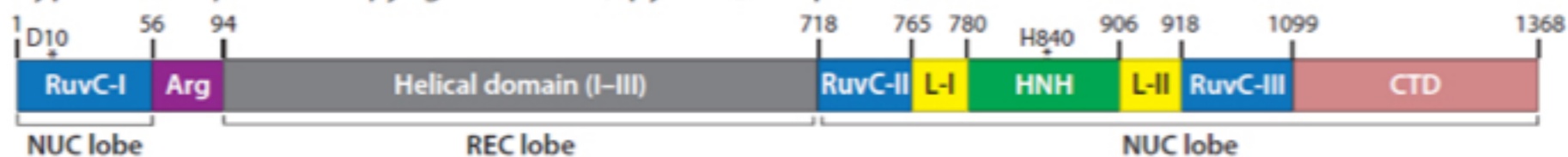
- ① RNA 结合结构域;
- ② a PAM-interacting site.
- ③ an  $\alpha$ -helical recognition lobe;
- ④ a nuclease lobe.

NHN: 负责切割与crRNA互补的链

RuvC: 负责非互补链的切割



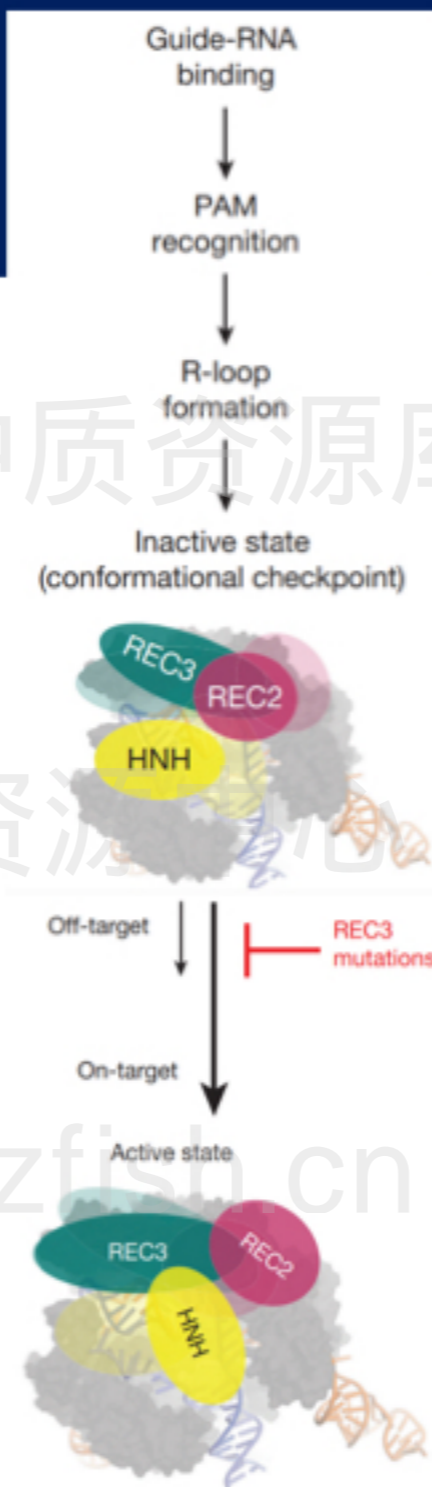
Type II-A *Streptococcus pyogenes* Cas9 (SpyCas9, or SpCas9)



### 3、基因编辑技术

#### Recognition (REC) lobe

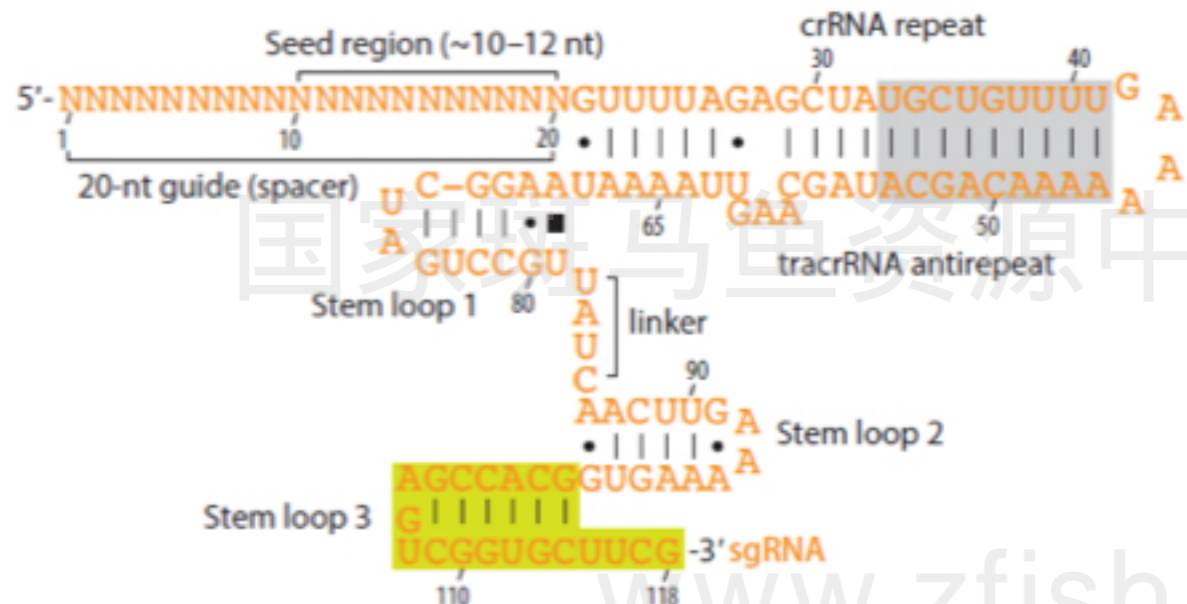
- REC3结构域
- REC2结构域



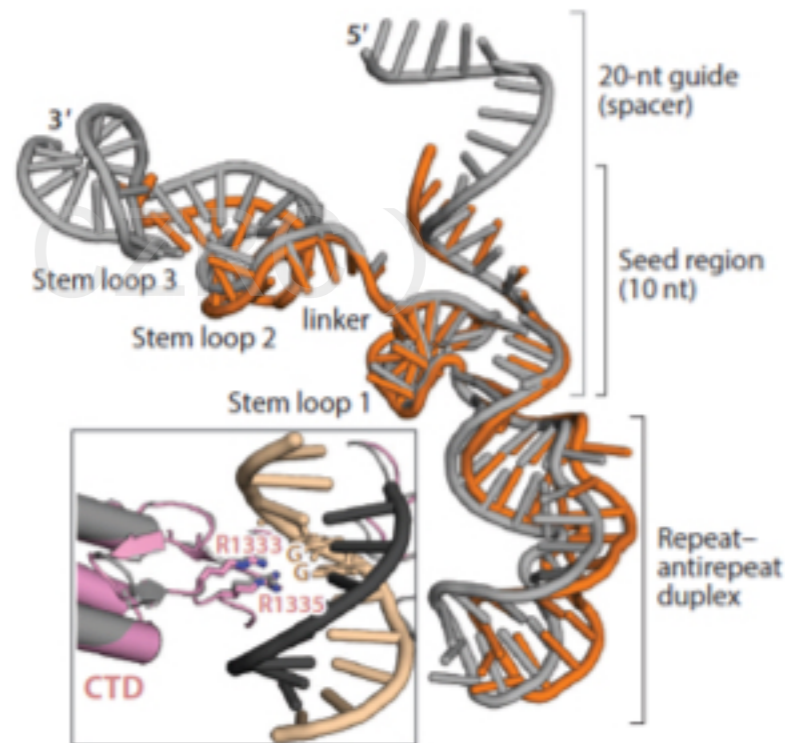
www.zfish.cn

# 3、基因编辑技术

- **Seed region:** gRNA 3'端10-12个nt, 决定了基因编辑的精确性



- sgRNA序列结构

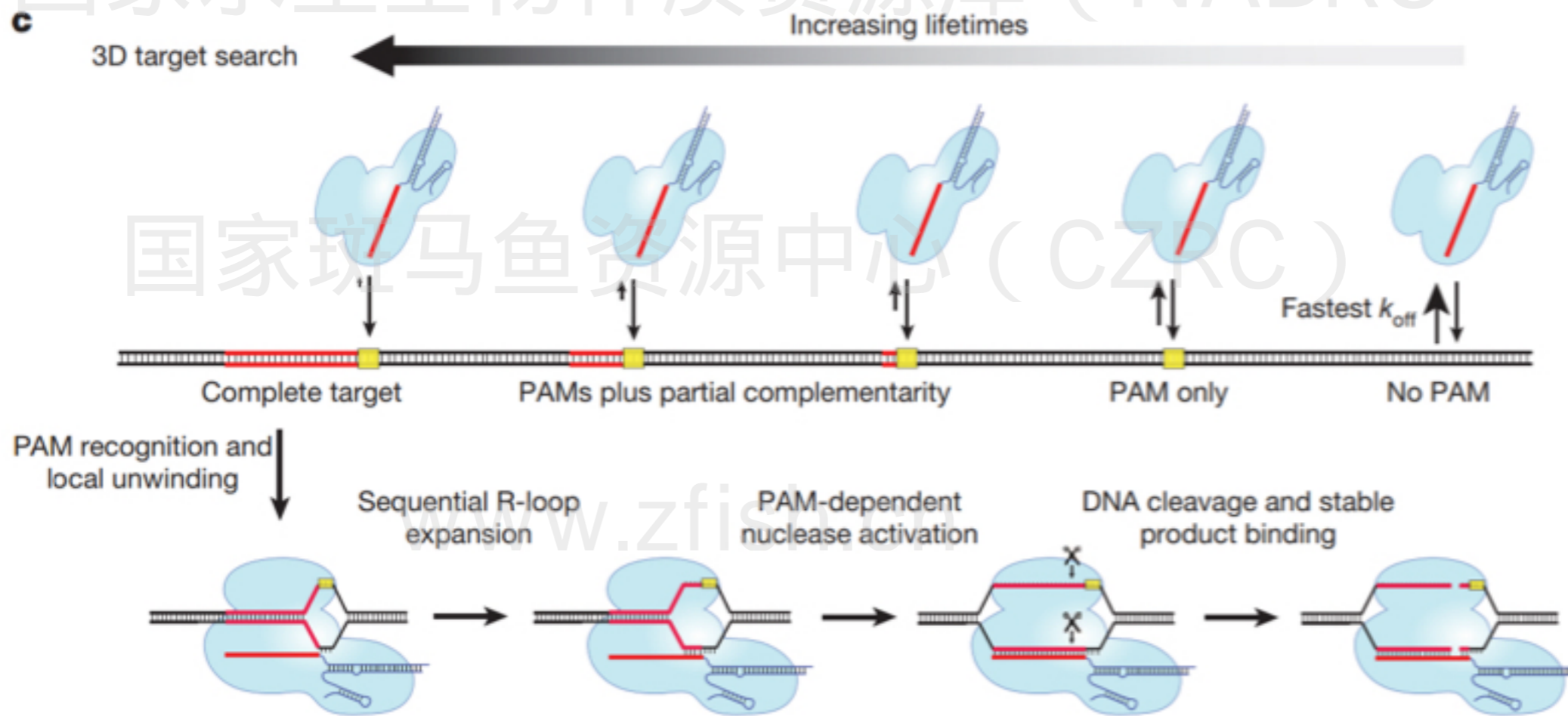


(Fuguo Jiang, et al., Annu. Rev. Biophys. 2017)



# 3、基因编辑技术

- **PAM区 (proto-spacer adjacent motif) & Seed region** 决定了sgRNA/Cas9复合物对靶位点的精确识别和切割



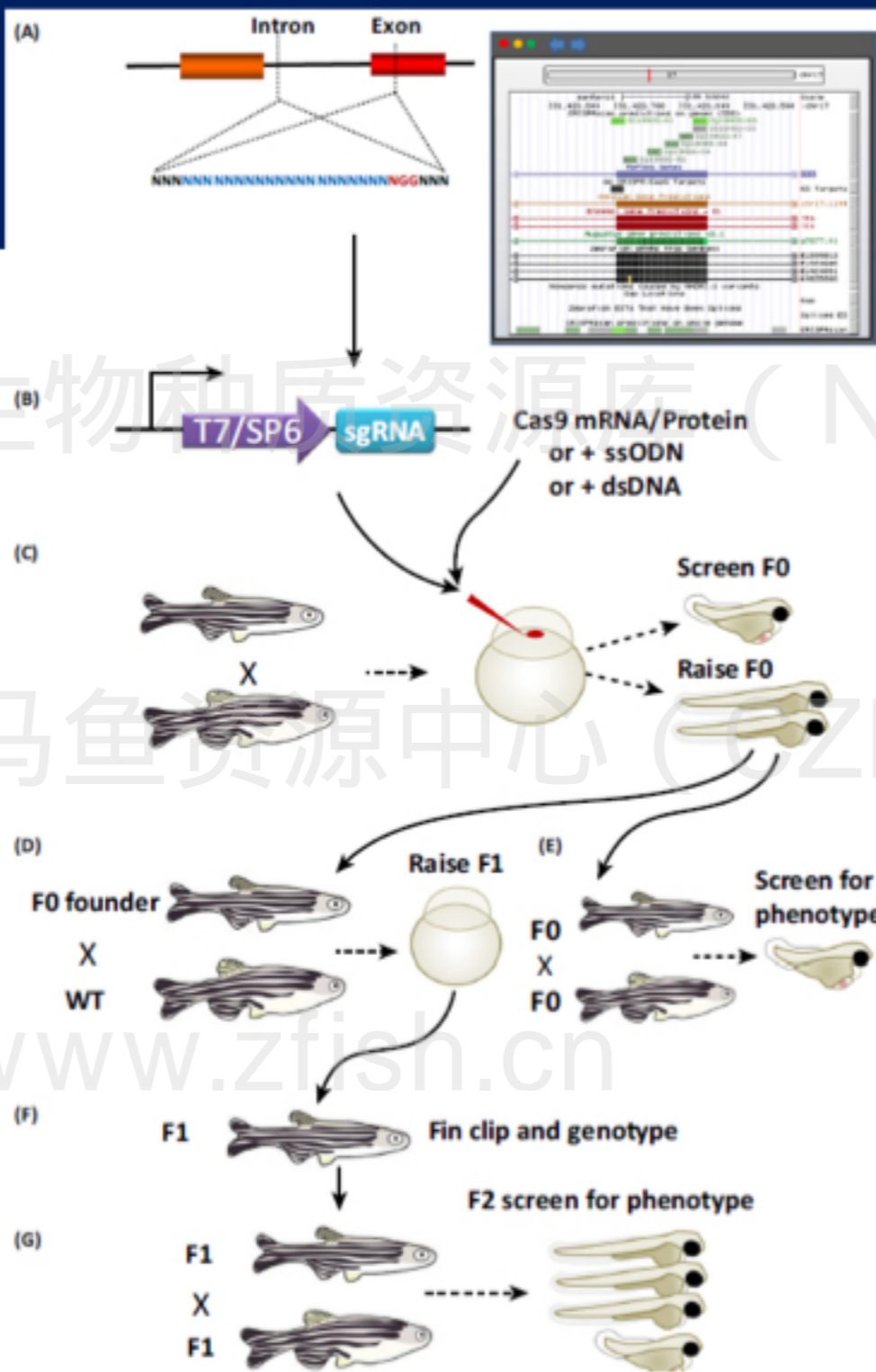
# 3、基因编辑技术

gRNA靶点设计, 合成gRNA和Cas9 mRNA

显微注射到斑马鱼胚胎中, 获取阳性F0代

F0代个体筛选, 通过与野生型侧交的方式进行, 将阳性胚胎培养为F1代

F1代个体筛选, 通过剪尾鳍测序的方式进行, 获取可稳定遗传的F1代个体



• 基于CRISPR/Cas9技术的基因敲除品系构建流程

# 3、基因编辑技术

## CRISPR/Cas9技术在斑马鱼研究中的应用

2013年，多家实验室成功使用CRISPR/Cas9技术对斑马鱼基因组进行精确的基因编辑。

nature  
biotechnology

Efficient genome editing in zebrafish using a CRISPR-Cas system

Woong Y Hwang<sup>1,7</sup>, Yanfang Fu<sup>2,3,7</sup>, Deepak Reyon<sup>1,3</sup>, Morgan L Maeder<sup>2,4</sup>, Shengdar Q Tsai<sup>1,3</sup>, Jeffrey D Sander<sup>2,3</sup>, Randall T Peterson<sup>1,5,6</sup>, J-R Joanna Yeh<sup>1,3</sup> & J Keith Joung<sup>2-4</sup>

Efficient multiplex biallelic zebrafish genome editing using a CRISPR nuclease system

Li-En Jao<sup>1</sup>, Susan R Wente<sup>1,2</sup>, and Wenbiao Chen<sup>1,2</sup>

<sup>1</sup>Department of Cell and Developmental Biology and <sup>2</sup>Department of Molecular Physiology and Biophysics, Vanderbilt University School of Medicine, Nashville, TN 37232-8240

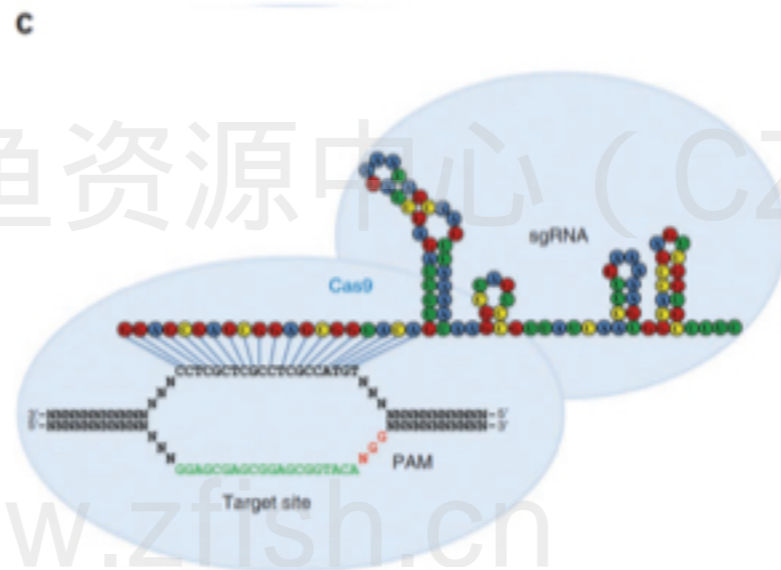
Edited by Igor B Dawid, The Eunice Kennedy Shriver National Institute of Child Health and Human Development, National Institutes of Health, Bethesda, MD, and approved July 12, 2013 (received for review May 3, 2013)

Cell Research (2013) 23:485-492.  
© 2013 IBCB, SIBS, CAS. All rights reserved 1001-0602/13  
www.nature.com/cr

Open  
ORIGINAL ARTICLE

Genome editing with RNA-guided Cas9 nuclease in Zebrafish embryos

Nannan Chang<sup>1,2,3</sup>, Changhong Sun<sup>1,2,4</sup>, Lu Gao<sup>1,3</sup>, Dan Zhu<sup>2</sup>, Xiufei Xu<sup>2</sup>, Xiaojun Zhu<sup>1,3</sup>, Jing-Wei Xiong<sup>1,3</sup>, Jianzhong Jeff Xi<sup>1,2</sup>



d

*tia11*  
Mutations in 17 out of 44 sequenced alleles

Sequence	Wild-type	Position
CCTGTGCTCCTCTGTTTTTAAGATATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	Wild-type	
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	+14 (-1, +15)	
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	+4 (-7, +11)	
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	+3 (-3, +6)	
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	-2	[+5]
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	-3 (-4, +1)	
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	-4	
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	-5	
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	-6	
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	-11	[+3]
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	-13	
CCTGTGCTCCTCTGTTTTTATAGTATGTCGGGAACCTCCTCAGGATGTTACGGAGCCCT	-23	

*gsk3b*

Mutations in 8 out of 16 sequenced alleles

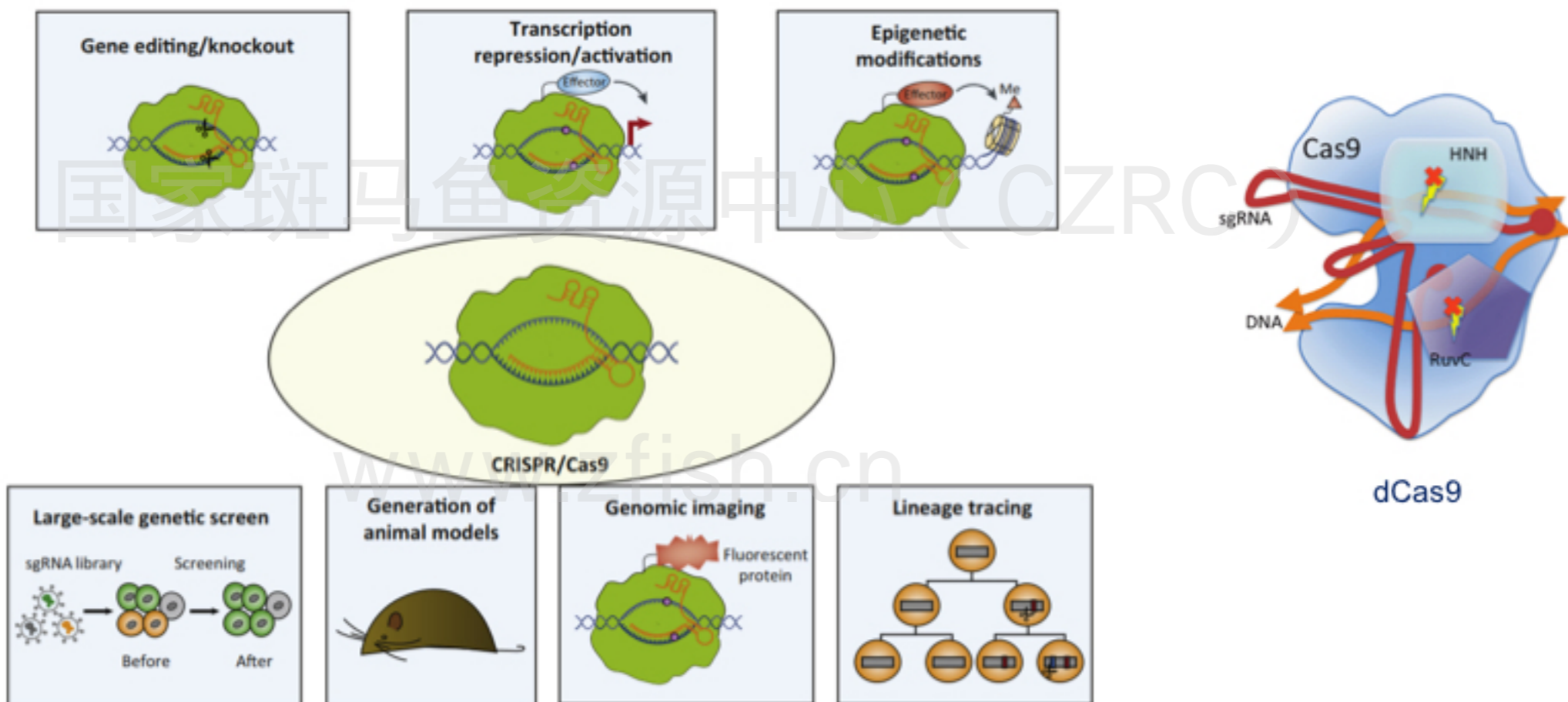
Sequence	Wild-type	Position
GTGGTGGGACTCCTGGACAGGGAAGCTGACCCCTCAGCTACACTGACACC	Wild-type	
GTGGTGGGACTCCTGGACAGGGAAGCTGACCCCTCAGCTACACTGACACC	+17 (-8, +25)	
GTGGTGGGACTCCTGGACAGGGAAGCTGACCCCTCAGCTACACTGACACC	+2 (-4, +6)	
GTGGTGGGACTCCTGGACAGGGAAGCTGACCCCTCAGCTACACTGACACC	+1 (-2, +3)	
GTGGTGGGACTCCTGGACAGGGAAGCTGACCCCTCAGCTACACTGACACC	+1 (-8, +9)	
GTGGTGGGACTCCTGGACAGGGAAGCTGACCCCTCAGCTACACTGACACC	-7	
GTGGTGGGACTCCTGGACAGGGAAGCTGACCCCTCAGCTACACTGACACC	-10	
GTGGTGGGACTCCTGGACAGGGAAGCTGACCCCTCAGCTACACTGACACC	-11	
GTGGTGGGACTCCTGGACAGGGAAGCTGACCCCTCAGCTACACTGACACC	-13	

- Targeted indel mutations induced by engineered sgRNA:Cas9 at the *tia11* and *gsk3b* genes.

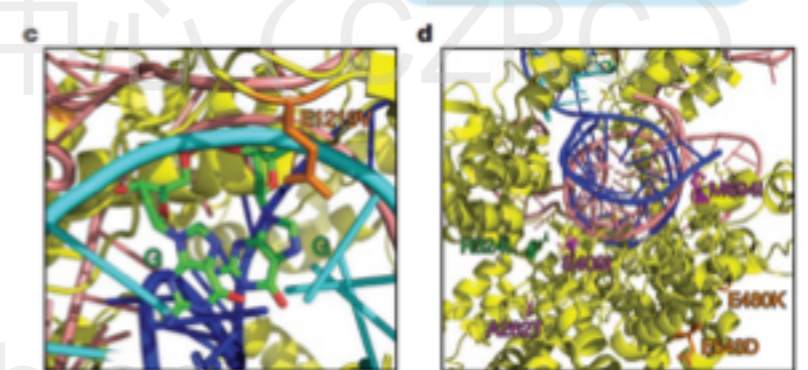
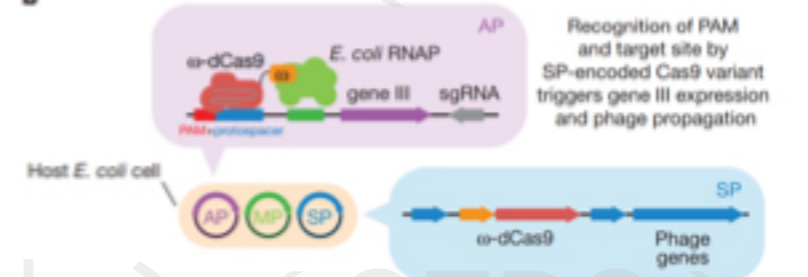
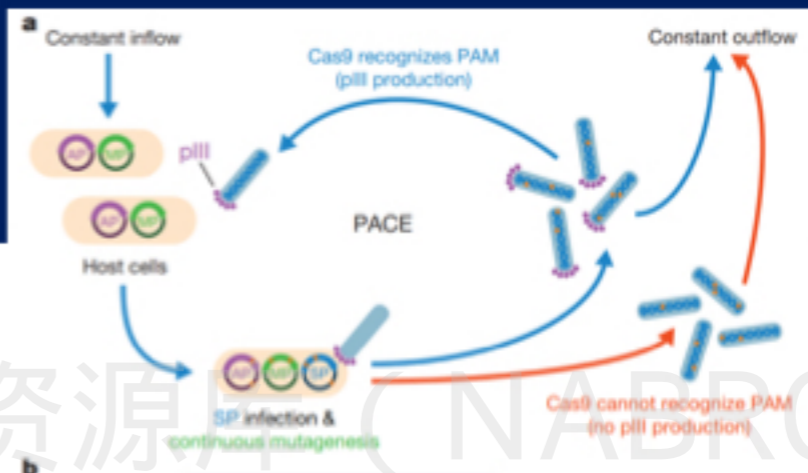
# 3、基因编辑技术

## CRISPR/Cas技术应用

应用于基因的敲除、敲入以及基因沉默、激活、表观遗传学、基因组成像等...



# 3、基因编辑技术



**e**

	S. pyogenes Cas9 amino acid									
	108	217	262	324	409	480	543	694	1219	
SpCas9	E	S	A	R	S	E	E	M	E	
xCas9-1.2	E	S	A	R	S	K	D	M	V	
xCas9-1.3	E	S	A	R	S	K	D	M	V	
xCas9-2.0	E	S	T	R	I	K	D	M	V	
xCas9-2.2	E	S	T	R	I	K	D	I	V	
xCas9-3.6	G	A	T	R	I	K	D	I	V	
xCas9-3.7	E	S	T	L	I	K	D	I	V	

**xCAS9: 适用范围扩大4倍, 特异性提升百倍**  
 可用于转录激活、DNA切割, 单碱基编辑



Article  
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**Evolved Cas9 variants with broad PAM compatibility and high DNA specificity**

Johnny H. Hu, Shannon M. Miller, Maarten H. Groot, Hee-yeon Song, Liwei Chen, Ning Sun, Christina M. Zerna, Xue Gao, Kelly S. Reed, Zhu-Li Li, David A. Liu

Nature  
 doi:10.1038/nature26155  
 Download Citation

Received: 27 January 2018  
 Accepted: 22 February 2018  
 Published online: 28 February 2018

	<b>spCAS9</b>	<b>xCAS9</b>
识别序列	1/16 (NGG)	1/4(NGN), GAA,GAT
转录激活		前者的数倍
DNA切割		较前者效率高
致病突变单碱基编辑 (C.G-T.A)	适用范围26%	适用范围73%
脱靶效应	少	前者的~1/100

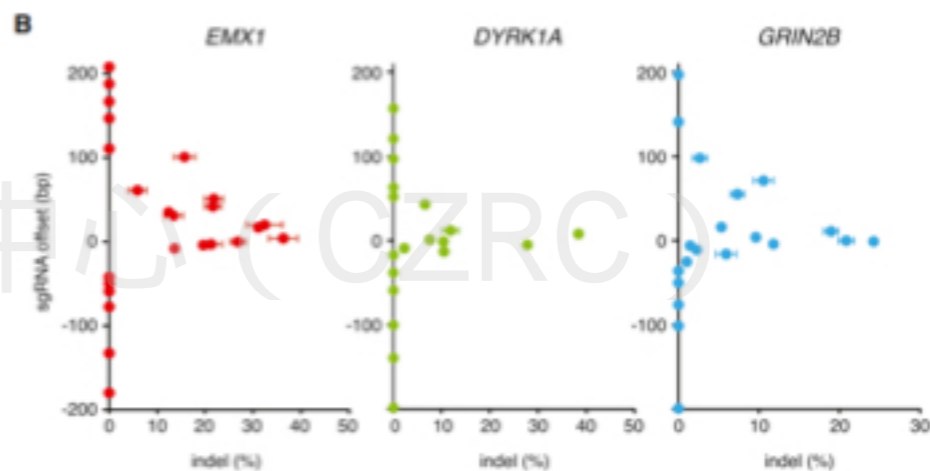
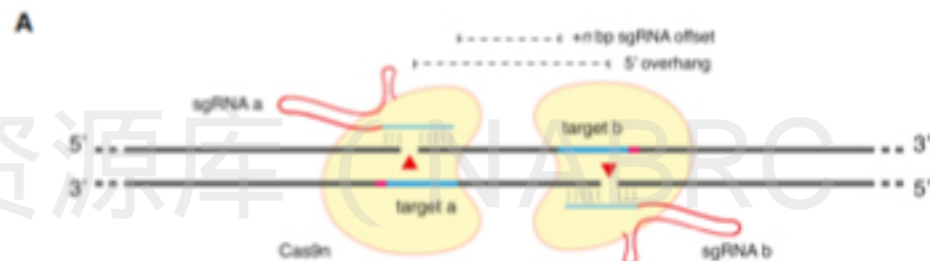
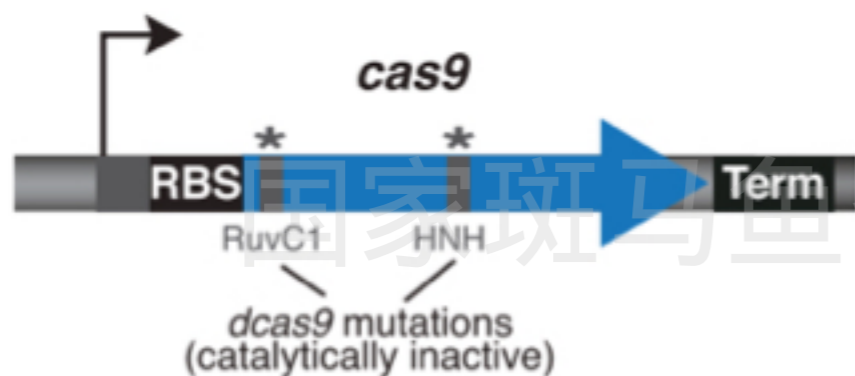
- PACE of Cas9 variants with broadened PAM compatibility.

# 3、基因编辑技术

## Cas9 Nickase (with paired gRNAs)

H840A in HNH or D10A in RuvC

特异性提升50-1500倍



## Resource

Cell

## Double Nicking by RNA-Guided CRISPR Cas9 for Enhanced Genome Editing Specificity

F. Ann Ran,<sup>1,2,3,4,5,11</sup> Patrick D. Hsu,<sup>1,2,3,4,5,11</sup> Chie-Yu Lin,<sup>1,2,3,4,6</sup> Jonathan S. Gootenberg,<sup>1,2,3,4</sup> Silvana Konermann,<sup>1,2,3,4</sup> Alexandro E. Trevino,<sup>1</sup> David A. Scott,<sup>1,2,3,4</sup> Azusa Inoue,<sup>7,8,9,10</sup> Shogo Matoba,<sup>7,8,9,10</sup> Yi Zhang,<sup>7,8,9,10</sup> and Feng Zhang<sup>1,2,3,4,\*</sup>

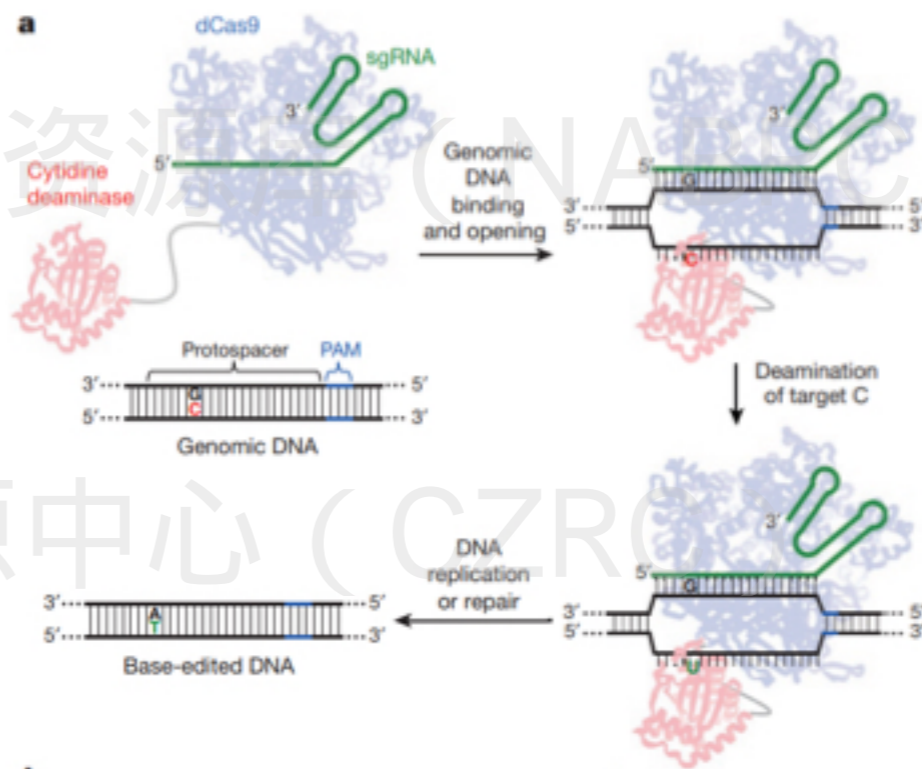
- Double Nicking Facilitates Efficient Genome Editing in Human Cells (Ran FA, et al. Cell. 2013. )

# 3、基因编辑技术

## Base Editing

2016年, David Liu团队将dCas9与胞苷脱氨酶相连, 实现C->T单碱基编辑

BE1: rat APOBEC1-XTEN-dCas9, 0.8%-7.7% in cells



LETTER

## Programmable editing of a target base in genomic DNA without double-stranded DNA cleavage

Alexis C. Komor<sup>1,2</sup>, Yongjoo B. Kim<sup>1,2</sup>, Michael S. Packer<sup>1,2</sup>, John A. Zuris<sup>1,2</sup> & David R. Liu<sup>1,2</sup>

www.zfish.cn

- First-generation base editor (BE1) mediates specific, guide RNA-programmed C→U conversion in vitro

# 3、基因编辑技术

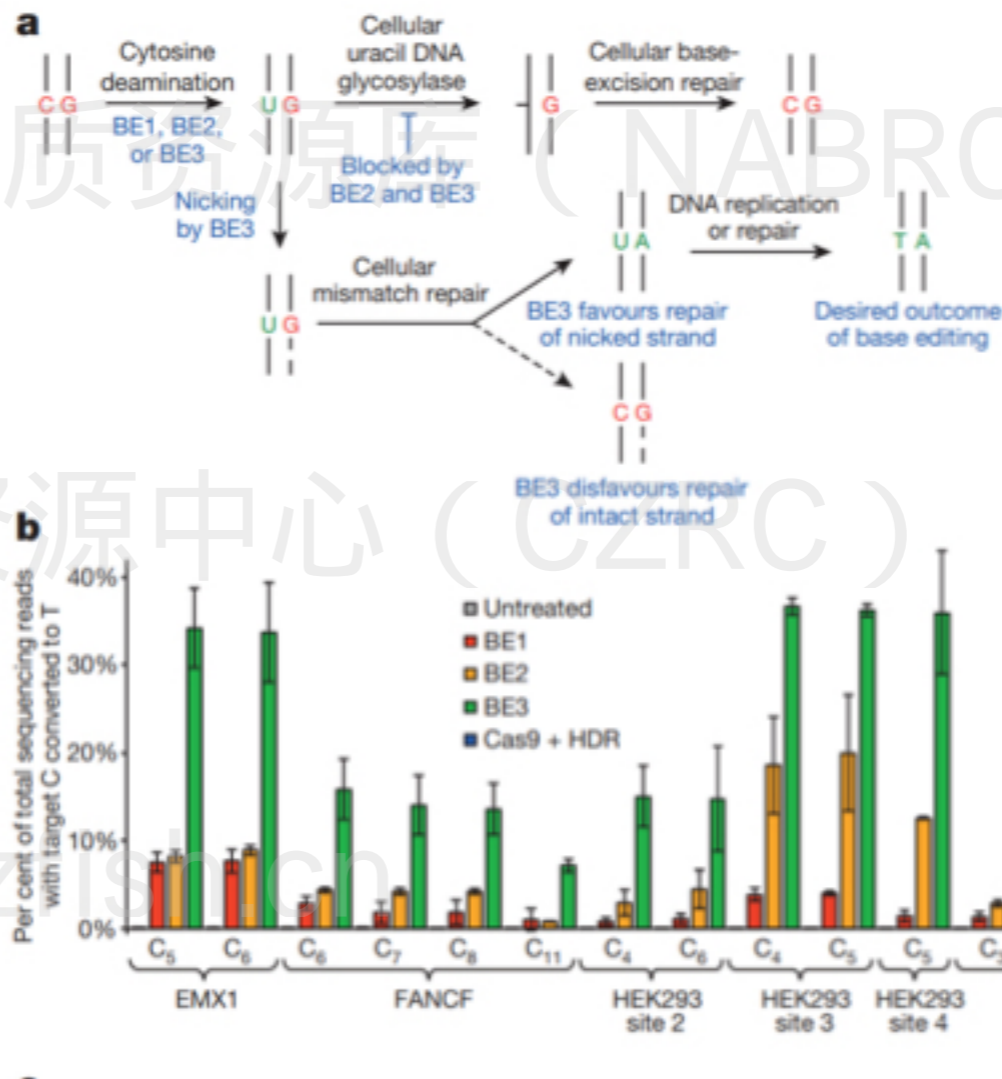
## Base Editing

BE3 APOBEC-XTEN-dCas9(A840H)-UGI

BE1: rat APOBEC1-XTEN-dCas9, 0.8%-7.7% in cells

BE2: APOBEC-XTEN-dCas9-UGI, 效率提高了3倍左右

BE3: APOBEC-XTEN-dCas9(A840H)-UGI, 效率又提高了2-6倍, 37% in cells  
Cas9 Nickase D10A

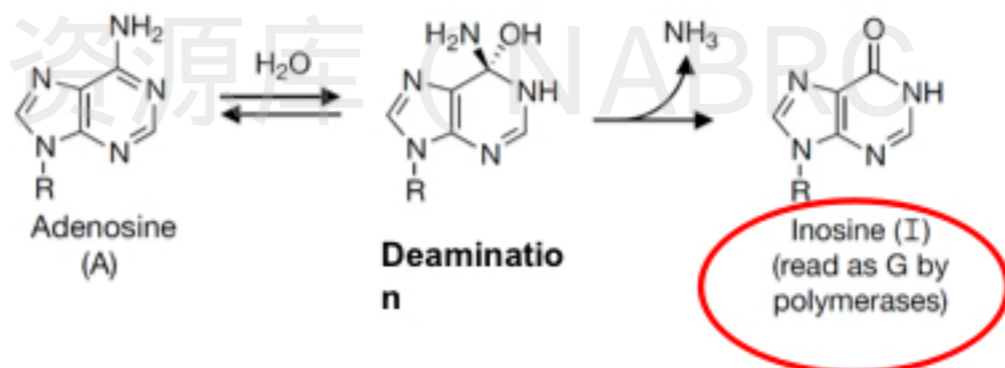
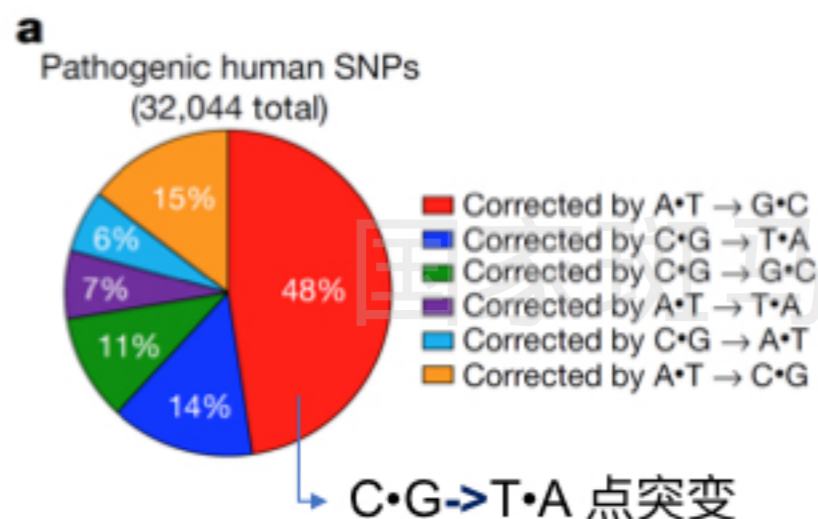




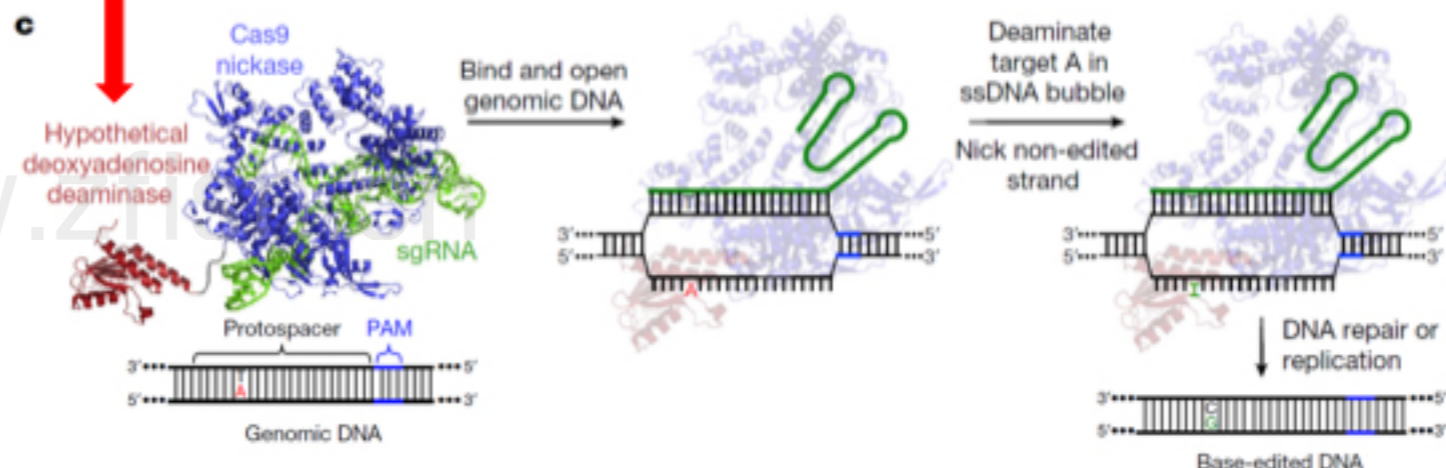
# 3、基因编辑技术

## Adenine Base Editors

2017年, 实现 A•T→G•C 碱基编辑



APOBEC-XTEN-dCas9(A840H)-UGI BE3  
**ecTadA**-XTEN-dCas9(A840H) ABE0.1



ARTICLE

## Programmable base editing of A•T to G•C in genomic DNA without DNA cleavage

Nicole M. Gaudelli<sup>1,2,3</sup>, Alexis C. Komor<sup>1,2,3</sup>†, Holly A. Rees<sup>1,2,3</sup>, Michael S. Packer<sup>1,2,3</sup>†, Ahmed H. Badran<sup>1,2,3</sup>, David I. Bryson<sup>1,2,3</sup>† & David R. Liu<sup>1,2,3</sup>

doi:10.1038/nature24644

• ABE-mediated A•T to G•C base editing strategy

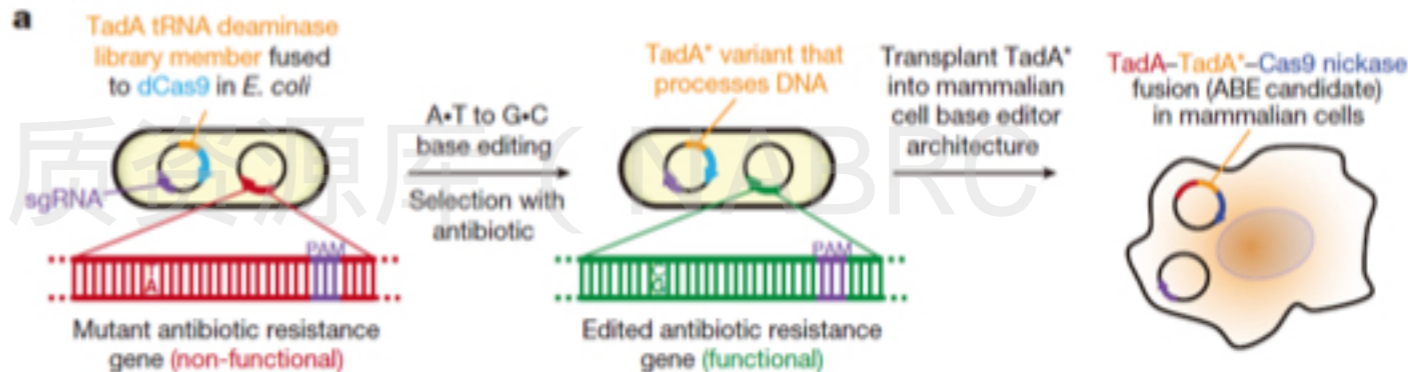
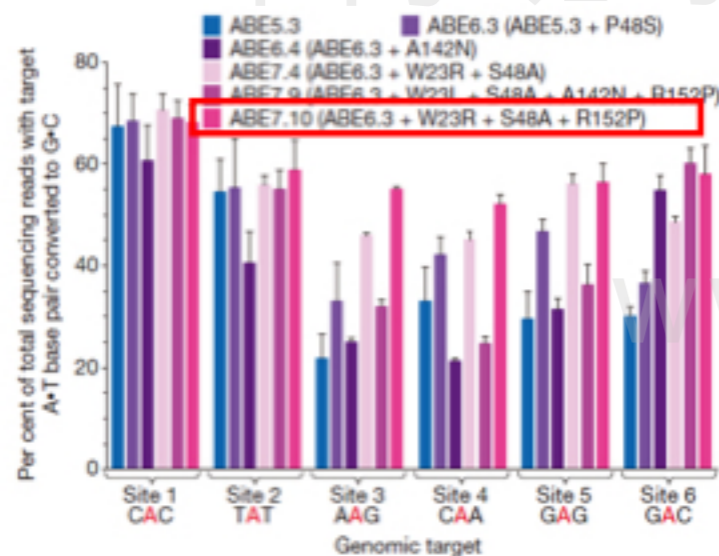
# 3、基因编辑技术

## Adenine Base Editors

wtTadA-linker1-TadA\*-linker2-nCas9-NLS ABE7.10

ABE1.2: TadA\*-XTEN-nCas9-NLS,  $3.2 \pm 0.88\%$  in cells

ABE7.10: wtTadA-linker1-TadA\*-linker2-nCas9-NLS,  $53 \pm 3.7\%$  in cells



**b**

*E. coli* TadA amino acid

	23	36	48	51	84	106	108	123	142	146	147	152	155	156	157	TadA state	Linker 1 length	Linker 2 length
ABE0.1	W	H	P	R	L	A	D	H	A	S	D	R	E	I	K	Monomer		16
ABE1.1	W	H	P	R	L	A	N	H	A	S	D	R	E	I	K	Monomer		16
ABE1.2	W	H	P	R	L	V	N	H	A	S	D	R	E	I	K	Monomer		16
ABE2.1	W	H	P	R	L	V	N	H	A	S	Y	R	V	I	K	Monomer		16
ABE2.9	W	H	P	R	L	V	N	H	A	S	Y	R	V	I	K	Homodimer	32	16
ABE2.10	W	H	P	R	L	V	N	H	A	S	Y	R	V	I	K	Heterodimer	32	16
ABE3.1	W	H	P	R	F	V	N	Y	A	S	Y	R	V	F	K	Homodimer	32	32
ABE4.3	W	H	P	R	F	V	N	Y	N	S	Y	R	V	F	K	Homodimer	32	32
ABE5.1	W	L	P	L	F	V	N	Y	A	C	Y	R	V	F	N	Homodimer	32	32
ABE5.3	W	L	P	L	F	V	N	Y	A	C	Y	R	V	F	N	Heterodimer	32	32
ABE6.3	W	L	S	L	F	V	N	Y	A	C	Y	R	V	F	N	Heterodimer	32	32
ABE6.4	W	L	S	L	F	V	N	Y	N	C	Y	R	V	F	N	Heterodimer	32	32
ABE7.4	R	L	A	L	F	V	N	Y	A	C	Y	R	V	F	N	Heterodimer	32	32
ABE7.8	L	L	A	L	F	V	N	Y	N	C	Y	R	V	F	N	Heterodimer	32	32
ABE7.9	L	L	A	L	F	V	N	Y	N	C	Y	P	V	F	N	Heterodimer	32	32
ABE7.10	R	L	A	L	F	V	N	Y	A	C	Y	P	V	F	N	Heterodimer	32	32

• Protein evolution and engineering of ABEs Gaudelli NM, et al. Nature. 2017)

# 3、基因编辑技术

## Minimization of spCas9

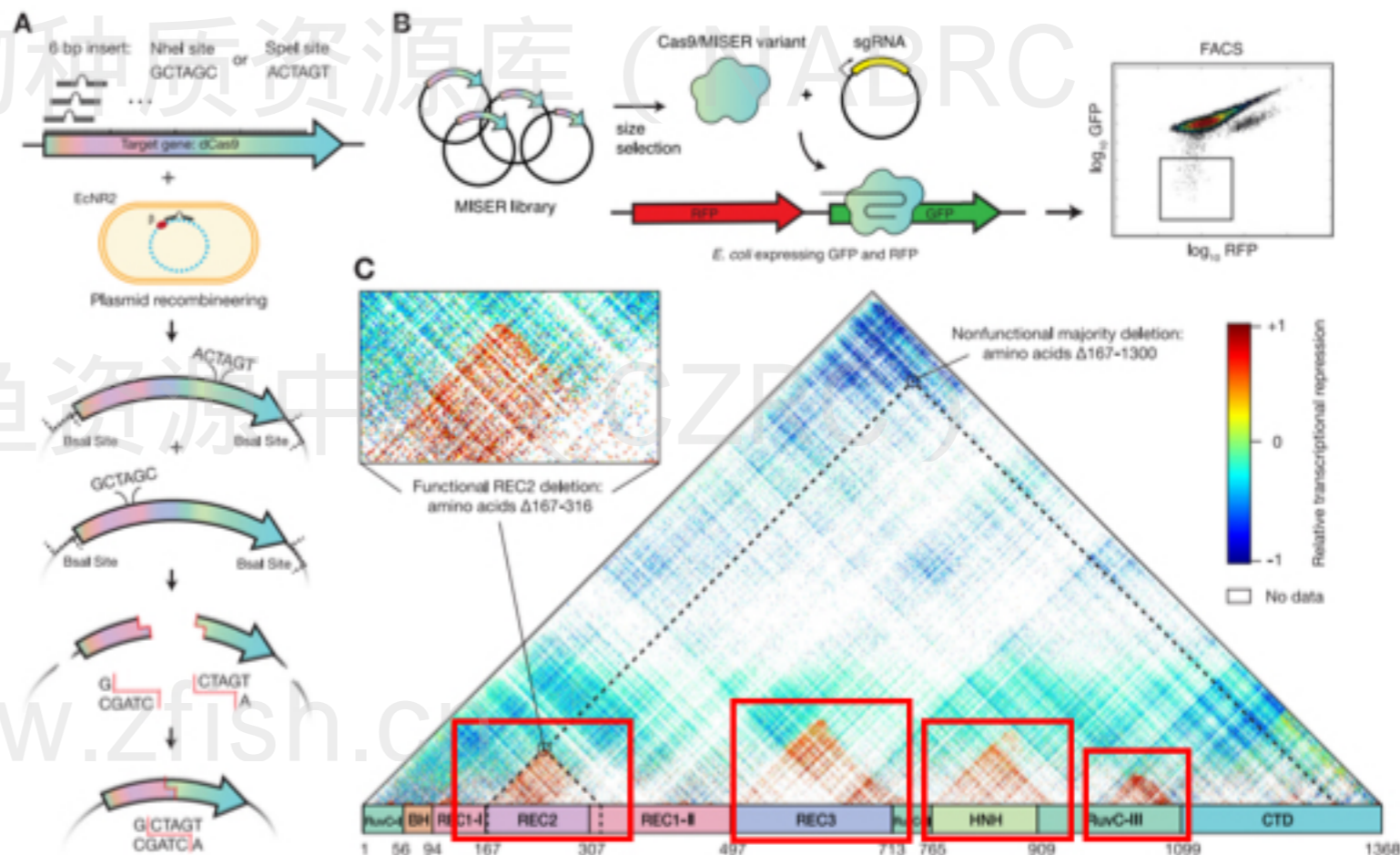
1368aa -> 874aa

ARTICLE

<https://doi.org/10.1038/s41467-021-25492-8> OPEN

Comprehensive deletion landscape of CRISPR-Cas9 identifies minimal RNA-guided DNA-binding modules

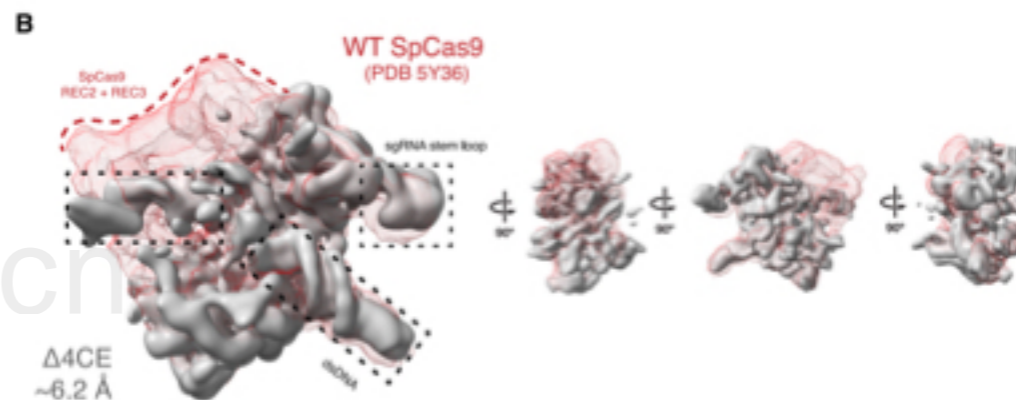
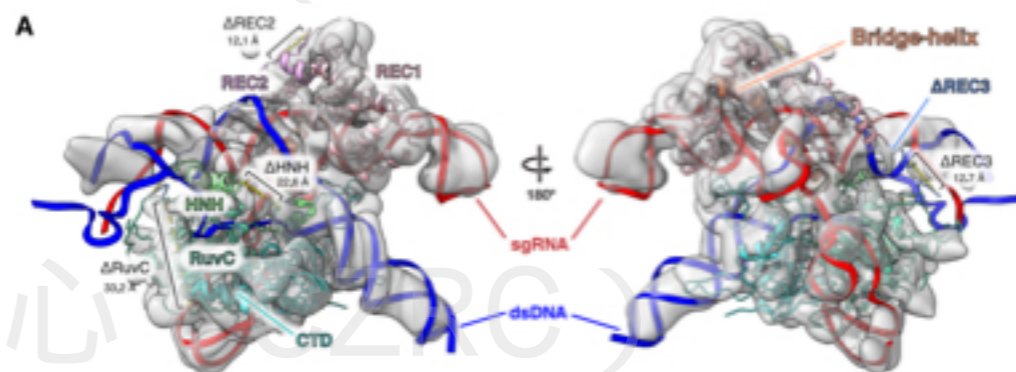
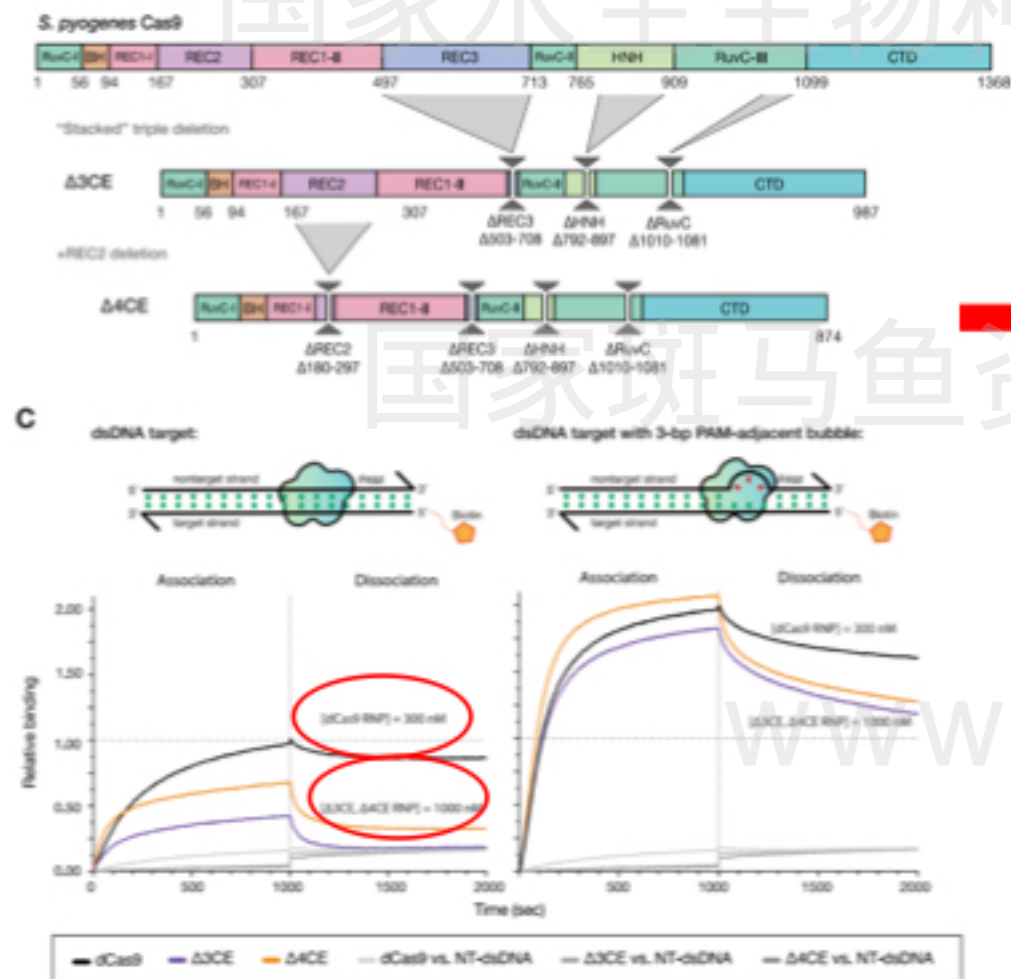
Arik Shams<sup>1,12</sup>, Sean A. Higgins<sup>1,2,3,12</sup>, Christof Fellmann<sup>1,4,5</sup>, Thomas G. Laughlin<sup>1,6</sup>, Benjamin L. Oakes<sup>1,2,3</sup>, Rachel Lew<sup>4</sup>, Shin Kim<sup>1,2</sup>, Maria Lukarska<sup>1,2</sup>, Madeline Arnold<sup>1</sup>, Brett T. Staahl<sup>1,2,3</sup>, Jennifer A. Doudna<sup>1,2,4,7,8,9,10,11</sup> & David F. Savage<sup>1,2,10</sup>



- Minimization by iterative size exclusion and recombination (**MISER**)

# 3、基因编辑技术

## Minimization of spCas9



- Density map of  $\Delta 4CE$  compared to WT SpCas9

(Shams A, et al. Nat Commun. 2021)

- Stacking multiple domain deletions on Cas9 results in defective DNA-binding activity

- 基因编辑技术的经历了四个发展阶段：经典同源重组、ZFNs、TALENs和CRISPR/Cas9技术。
- 三代基因编辑技术能够对基因组进行精确编辑。ZFNs和TALENs技术是通过氨基酸残基来特异性识别DNA碱基，CRISPR/Cas9技术是通过一小段RNA分子以碱基互补配对的方式来特异性识别DNA碱基，实现对基因组的特异性编辑。
- 三代基因编辑技术造成靶向序列DNA双链断裂，通过同源重组修复（HR）、非同源末端连接（NHEJ）以及微同源介导的末端连接/单链退火修复（MMEJ/SSA）机制进行DNA的损伤修复。
- 通过NHEJ、MMEJ/SSA途径进行修复，将造成靶向序列处有碱基插入或缺失，导致基因的敲除；通过HR途径在提供外源DNA模版的条件下，靶向序列处将得到精确的修复或外源基因的敲入。
- 利用CRISPR/Cas9技术可以实现多种多样的基因调控方式，最基本的就是基因敲除和敲入，其次是使用内切酶活性失活的dCas9对基因组的特定定位点进行基因调控和递送各种功能因子。

国家水生生物种质资源库 (NABRC)  
**本讲内容完毕**

**欢迎交流!**  
国家斑马鱼资源中心 (CZRC)



中国斑马鱼信息中心